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Patentanmeldung Nr.

Patent application No. Demande de brevet n°

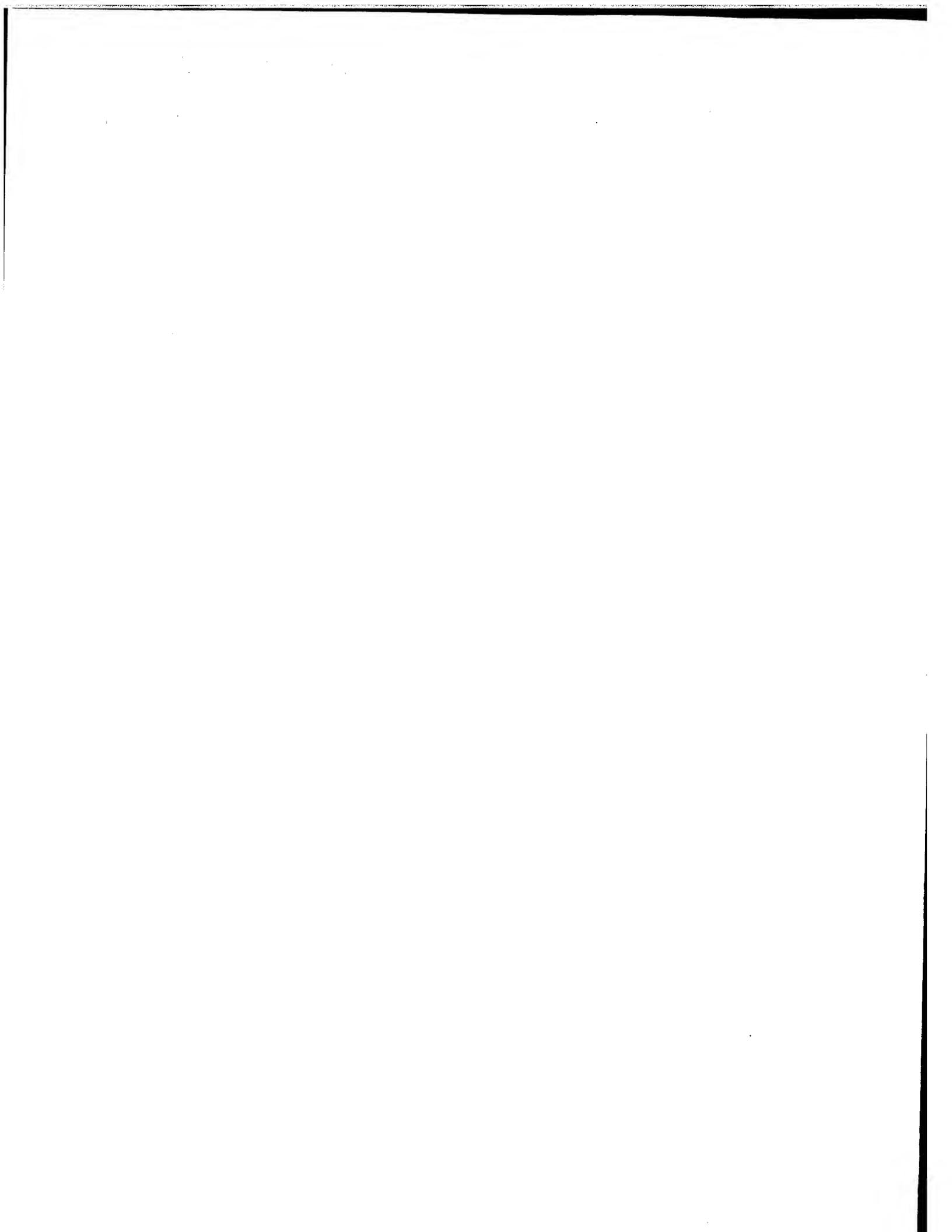
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For the President of the European Patent Office

Le Président de l'Office européen des brevets p.o.

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Ornithobacterium rhinotracheale subunit vaccines

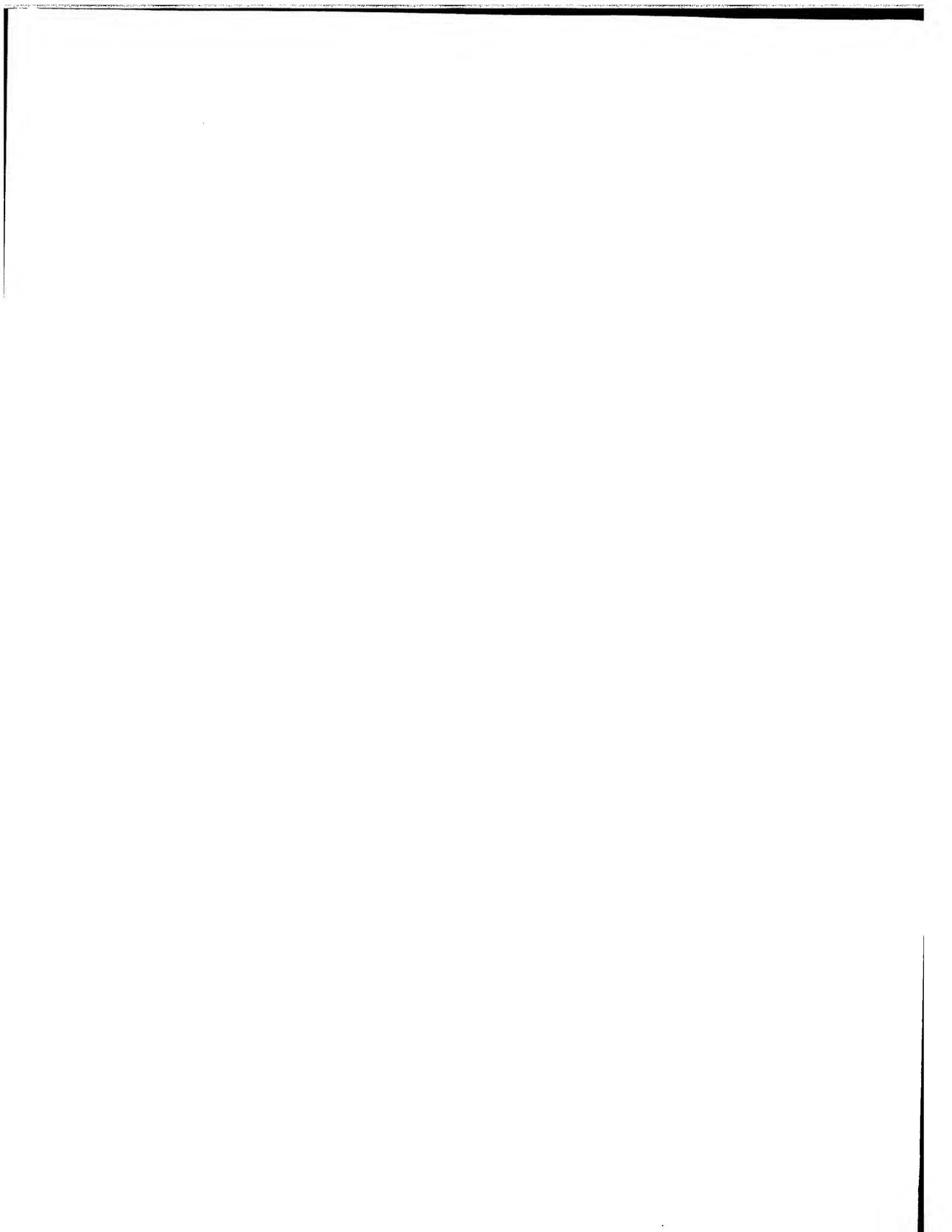
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Ornithobacterium rhinotracheale subunit vaccines.

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The present invention relates to nucleic acids encoding *Ornithobacterium rhinotracheale* proteins, to DNA fragments, recombinant DNA molecules, live recombinant carriers and host cells comprising such nucleic acids, to *Ornithobacterium rhinotracheale* proteins, to antibodies against such proteins, to such proteins for use in vaccines, to the use of such proteins in the manufacturing of such vaccines, to vaccines comprising such nucleic acids, DNA fragments, recombinant DNA molecules, live recombinant carriers, host cells, proteins or antibodies against such proteins, and to methods for the preparation of such vaccines.

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Ornithobacterium rhinotracheale is a relatively recently discovered bacterium that is found more and more frequently in poultry farms, and in wild birds. Especially animals in commercial chicken farms, turkey farms and duck farms are frequently infected.

In commercial poultry, infection is associated with respiratory diseases: airsacculitis and pneumonia are the most common features of infection with Ornithobacterium rhinotracheale. These signs can be induced by aerosol in intra-tracheal or intra-thoracic administration of the organism and are aggravated by other factors such as respiratory viruses, bacteria or sub-optimal housing conditions. Osteitis, meningitis and joint-infections which can be induced by intravenous application have been associated with Ornithobacterium rhinotracheale. The infection can be transmitted horizontally, as well as vertically through eggs, which probably accounts for its rapid and worldwide spread. An extensive review of Ornithobacterium rhinotracheale has been given by van Empel, P.C.M. ad Hafez, H.M. in Avian Pathology 28:217-227 (1999). European Patent EP0.625.190 relates to both the Ornithobacterium rhinotracheale.

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Serological research has revealed that *Ornithobacterium rhinotracheale* strains may have different serotypes, to a certain degree depending on the geographic origin of the strain and the host animal from which they were isolated. At this moment, eighteen different serotypes are found.

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Therapeutic treatment of the disease can be difficult because acquired resistance against the regular antibiotics is very common within the genus. Moreover, there is an increasing reluctance against the use of antibiotics in food animals for both public health- and environmental reasons.

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Vaccination offers an alternative for therapeutic treatment with antibiotics, but up till now, only vaccination with live attenuated vaccines and inactivated whole cell vaccines was possible.

- The success of live attenuated vaccines specifically for *Ornithobacterium rhinotracheale* depends highly on the right balance between attenuation and triggering of the immune system. Inactivated whole cell vaccines are basically safe and therefore, from a safety point of view would seem the preferred type of vaccine.
- Inactivated whole cell vaccines however need to be given in a higher dose compared to live attenuated vaccines. As a general rule, most of the proteins present in a bacterium play no role in the triggering of the immune system, i.e. they are not relevant immunogens. This means that, in the case of inactivated whole cell vaccines, in order to provide humans or animals with a sufficient level of relevant immunogens a lot of non-protective material is additionally and unavoidably administered. This is not a desirable situation.

The use of subunit vaccines could overcome this problem, and would therefore be highly preferred, but currently no immunogenic subunit vaccines are known in the art for combating *Ornithobacterium rhinotracheale*.

Moreover, although live attenuated vaccines and inactivated whole cell preparations are known to provide a certain level of cross-protection against all *Ornithobacterium* rhinotracheale strains, subunit vaccines might or might not induce cross-reactivity.

The present invention aims at providing for the first time vaccines that are based upon Ornithobacterium rhinotracheale subunits that do induce cross-reactivity.

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This objective is reached by providing eight novel *Ornithobacterium rhinotracheale* proteins that surprisingly play an important role in triggering a protective immune response, and by providing vaccines comprising one or more of these novel immunogenic proteins.

Even more surprisingly, these eight novel proteins were found no only to induce a protective homologous immune response, but to also induce a protective cross-reactive immune response.

A homologous immune response is a response against strains of the same serotype, whereas a cross-reactive immune response is a response against both serologically homologous and heterologous strains.

The first novel protein, Or01, having a molecular weight of 59.8 kD is encoded by a nucleic acid having a nucleotide sequence as depicted in SEQ ID NO: 1.

It is well-known in the art, that many different nucleotide sequences can encode one and the same protein. This phenomenon is commonly known as wobble in the second and especially the third base of each triplet encoding an amino acid. This phenomenon can result in a heterology of about 20-30% for two nucleotide sequences still encoding the same protein. Therefore, two nucleic acids having a nucleotide sequence homology of about 80 % can still encode one and the same protein.

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Thus, one embodiment relates to a nucleic acid encoding a 59.8 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein wherein said nucleic acid or said part thereof has at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO:

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The molecular weight of the protein (and the seven other proteins) is determined on the basis of the molecular weight of the amino acids as given in the amino acid sequence.

Preferably, a nucleic acid according to the invention encoding this 59.8 kD *Ornithobacterium* rhinotracheale protein or a part of that nucleic acid that encodes an immunogenic fragment of that protein has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO:

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Even more preferred is a homology level of 98 %, 99 % or even 100 %.

The level of nucleotide homology can be determined with the computer program "BLAST 2 SEQUENCES" by selecting sub-program: "BLASTN" that can be found at

30 www.ncbi.nlm.nih.gov/blast/bl2seq/bl2.html.

A reference for this program is Tatiana A. Tatusova, Thomas L. Madden FEMS Microbiol. Letters 174: 247-250 (1999). Parameters used are the default parameters: Reward for a match: +1. Penalty for a mismatch: -2. Open gap: 5. Extension gap: 2. Gap x_dropoff: 50.

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Another approach for deciding if a certain nucleic acid sequence is or is not a nucleic acid sequence according to the invention relates to the question if that certain nucleic acid sequence does hybridize under stringent conditions to the nucleotide sequence as depicted in SEQ ID NO: 1 (or in SEQ ID NO: 3, 5,7, 9, 11, 13 or 15, see below).

If a nucleic acid sequence hybridizes under stringent conditions to the nucleotide sequence as depicted in SEQ ID NO: 1, or of course as depicted in SEQ ID NO: 3, 5,7, 9, 11, 13 and 15, it is considered to be a nucleic acid sequence according to the invention.

The definition of stringent conditions follows from the formula of Meinkoth and Wahl (1984. Hybridization of nucleic acids immobilized on solid supports. Anal. Biochem. 138: 267-284.).

Tm = $[81.5^{\circ}C + 16.6(\log M) + 0.41(\%GC) - 0.61(\%formamide) - 500/L] - 1^{\circ}C/1\%mismatch$

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In this formula, M is molarity of monovalent cations; %GC is the percentage of guanosine and cytosine nucleotides in the DNA; L is the length of the hybrid in base pairs.

Stringent conditions are those conditions under which nucleic acid sequences or fragments thereof still hybridize, if they have a mismatch of 20 % at the most, preferably 10%, more preferably 8, 6, 5, 4,3, 2, 1 or 0% in that order or preference, to the nucleic acid sequence as depicted in any of the SEQ ID NO: 1, 3, 5,7, 9, 11, 13 or 15.

Another embodiment relates to a nucleic acid encoding a 58.2 kD Ornithobacterium rhinotracheale protein Or02, or a part of said nucleic acid that encodes an immunogenic fragment of said protein wherein said nucleic acid or said part thereof has at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 3.

Preferably, a nucleic acid according to the invention encoding this 58.2 kD Ornithobacterium rhinotracheale protein or a part of that nucleic acid that encodes an immunogenic fragment of that protein has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 3.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

Still another embodiment relates to a nucleic acid encoding a 46.0 kD Ornithobacterium rhinotracheale protein Or03 or a part of said nucleic acid that encodes an immunogenic fragment of said protein wherein said nucleic acid or said part thereof has at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 5.

Preferably, a nucleic acid according to the invention encoding this 46.0 kD *Ornithobacterium* rhinotracheale protein or a part of that nucleic acid that encodes an immunogenic fragment of that protein has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium* rhinotracheale protein gene as depicted in SEQ ID NO: 5.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

Again another embodiment relates to a nucleic acid encoding a 37.2 kD Ornithobacterium rhinotracheale protein Or04 or a part of said nucleic acid that encodes an immunogenic fragment of said protein wherein said nucleic acid or said part thereof has at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 7.

Preferably, a nucleic acid according to the invention encoding this 37.2 kD *Ornithobacterium* rhinotracheale protein or a part of that nucleic acid that encodes an immunogenic fragment of that protein has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO:

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Even more preferred is a homology level of 98 %, 99 % or even 100 %.

Another embodiment relates to a nucleic acid encoding a 45.6 kD Ornithobacterium rhinotracheale protein Or11 or a part of said nucleic acid that encodes an immunogenic fragment of said protein wherein said nucleic acid or said part thereof has at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 9.

Preferably, a nucleic acid according to the invention encoding this 45.6 kD Ornithobacterium rhinotracheale protein or a part of that nucleic acid that encodes an immunogenic fragment of that protein has at least 85 %, preferably 90 %, more preferably 95 % homology with the

nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 9.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

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Again another embodiment relates to a nucleic acid encoding a 42.2 kD *Ornithobacterium rhinotracheale* protein Or77 or a part of said nucleic acid that encodes an immunogenic fragment of said protein wherein said nucleic acid or said part thereof has at least 80 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 11.

Preferably, a nucleic acid according to the invention encoding this 42.2 kD Ornithobacterium rhinotracheale protein or a part of that nucleic acid that encodes an immunogenic fragment of that protein has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 11.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

Also another embodiment relates to a nucleic acid encoding a 34.0 kD Ornithobacterium rhinotracheale protein Or98A or a part of said nucleic acid that encodes an immunogenic fragment of said protein wherein said nucleic acid or said part thereof has at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 13.

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Preferably, a nucleic acid according to the invention encoding this 34.0 kD Ornithobacterium rhinotracheale protein or a part of that nucleic acid that encodes an immunogenic fragment of that protein has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO:

30 13.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

Another embodiment relates to a nucleic acid encoding a 32.9 kD Ornithobacterium

rhinotracheale protein Or98B or a part of said nucleic acid that encodes an immunogenic fragment of said protein wherein said nucleic acid or said part thereof has at least 80 %

homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 15.

Preferably, a nucleic acid according to the invention encoding this 32.9 kD *Ornithobacterium* rhinotracheale protein or a part of that nucleic acid that encodes an immunogenic fragment of that protein has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 15.

10 Even more preferred is a homology level of 98 %, 99 % or even 100 %.

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Nucleotide sequences that are complementary to the sequence depicted in SEQ ID NO 1, 3, 5, 7, 9, 11, 13 or 15 or nucleotide sequences that comprise tandem arrays of the sequences according to the invention are also within the scope of the invention.

Since the present invention discloses nucleic acids encoding 8 novel Ornithobacterium rhinotracheale proteins, it is now for the first time possible to obtain these proteins in significant quantities. This can e.g. be done by using expression systems to express the whole or parts of a gene encoding the protein or an immunogenic fragment thereof.

Therefore, in a preferred form of this embodiment, the invention relates to DNA fragments comprising a nucleic acid according to the invention. A DNA fragment is a stretch of nucleotides that functions as a carrier for a nucleic acid according to the invention. Such DNA fragments can e.g. be plasmids, into which a nucleic acid according to the invention is cloned. Such DNA fragments are e.g. useful for enhancing the amount of DNA for use as a primer and for expression of a nucleic acid according to the invention, as described below.

An essential requirement for the expression of the nucleic acid is an adequate promoter functionally linked to the nucleic acid, so that the nucleic acid is under the control of the promoter. It is obvious to those skilled in the art that the choice of a promoter extends to any eukaryotic, prokaryotic or viral promoter capable of directing gene transcription in cells used as host cells for protein expression.

Therefore, a more preferred form of this embodiment relates to a recombinant DNA molecule comprising a DNA fragment and/or a nucleic acid according to the invention wherein the nucleic acid according to the invention is placed under the control of a functionally linked promoter. This can be obtained by means of e.g. standard molecular biology techniques.

(Maniatis/Sambrook (Sambrook, J. Molecular cloning: a laboratory manual, 1989. ISBN 0-87969-309-6).

Functionally linked promoters are promoters that are capable of controlling the transcription of the nucleic acids to which they are linked.

- Such a promoter can be the native promoter of the novel gene, i.e. the promoter that is involved in the transcription of the nucleic acid encoding a protein according to the invention, or another promoter of *Ornithobacterium rhinotracheale*, provided that that promoter is functional in the cell used for expression. It can also be a heterologous promoter. When the host cells are bacteria, useful expression control sequences which may be used include the
- Trp promoter and operator (Goeddel, et al., Nucl. Acids Res., 8, 4057, 1980); the lac promoter and operator (Chang, et al., Nature, 275, 615, 1978); the outer membrane protein promoter (Nakamura, K. and Inouge, M., EMBO J., 1, 771-775, 1982); the bacteriophage lambda promoters and operators (Remaut, E. et al., Nucl. Acids Res., 11, 4677-4688, 1983); the α-amylase (B. subtilis) promoter and operator, termination sequences and other expression enhancement and control sequences compatible with the selected host cell.
 - When the host cell is yeast, useful expression control sequences include, e.g., α-mating factor. For insect cells the polyhedrin or p10 promoters of baculoviruses can be used (Smith, G.E. et al., Mol. Cell. Biol. 3, 2156-65, 1983). When the host cell is of vertebrate origin illustrative useful expression control sequences include the (human) cytomegalovirus immediate early promoter (Seed, B. et al., Nature 329, 840-842, 1987; Fynan, E.F. et al., PNAS 90, 11478-11482 1993; Illmer I.B. et al., Spinger 250, 1745, 1749, 1993; Fynan, E.F. et al., PNAS 90, 11478-
- promoter (Seed, B. et al., Nature 329, 840-842, 1987; Fynan, E.F. et al., PNAS 90, 11478-11482,1993; Ulmer, J.B. et al., Science 259, 1745-1748, 1993), Rous sarcoma virus LTR (RSV, Gorman, C.M. et al., PNAS 79, 6777-6781, 1982; Fynan et al., supra; Ulmer et al., supra), the MPSV LTR (Stacey et al., J. Virology 50, 725-732, 1984), SV40 immediate early promoter (Sprague J. et al., J. Virology 45, 773, 1983), the SV-40 promoter (Berman, P.W. et al., Science, 222, 524-527, 1983), the metallothionein promoter (Brinster, R.L. et al., Nature
- 296, 39-42, 1982), the heat shock promoter (Voellmy et al., Proc. Natl. Acad. Sci. USA, 82, 4949-53, 1985), the major late promoter of Ad2 and the β-actin promoter (Tang et al., Nature 356, 152-154, 1992). The regulatory sequences may also include terminator and polyadenylation sequences. Amongst the sequences that can be used are the well known bovine growth hormone poly-adenylation sequence, the SV40 poly-adenylation sequence, the human cytomegalovirus (hCMV) terminator and poly-adenylation sequences.

Bacterial, yeast, fungal, insect and vertebrate cell expression systems are very frequently used systems. Such systems are well-known in the art and generally available, e.g. commercially through Clontech Laboratories, Inc. 4030 Fabian Way, Palo Alto, California 94303-4607, USA. Next to these expression systems, parasite-based expression systems are attractive

expression systems. Such systems are e.g. described in the French Patent Application with Publication number 2 714 074, and in US NTIS Publication No US 08/043109 (Hoffman, S. and Rogers, W.: Public. Date 1 December 1993).

- An even more preferred form of this embodiment of the invention relates to Live
 Recombinant Carriers (LRCs) comprising a nucleic acid encoding an Ornithobacterium
 rhinotracheale protein or an immunogenic fragment thereof according to the invention, a
 DNA fragment according to the invention or a recombinant DNA molecule according to the
 invention. These LRCs are micro-organisms or viruses in which additional genetic
 information, in this case a nucleic acid encoding an Ornithobacterium rhinotracheale protein
 or an immunogenic fragment thereof, a DNA fragment or a recombinant DNA molecule
 according to the invention has been cloned. Chickens infected with such LRCs will produce
 an immunological response not only against the immunogens of the carrier, but also against
 the immunogenic parts of the protein(s) for which the genetic code is additionally cloned into
 the LRC, e.g. an Ornithobacterium rhinotracheale protein gene according to the invention.
 - As an example of bacterial LRCs, attenuated Salmonella strains known in the art can very attractively be used.
- Also, live recombinant carrier parasites have i.a. been described by Vermeulen, A. N. (Int. 20 Journ. Parasitol. 28: 1121-1130 (1998)).
 - Furthermore, LRC viruses may be used as a way of transporting the nucleic acid into a target cell. Live recombinant carrier viruses are also called vector viruses. Viruses often used as vectors are Vaccinia viruses (Panicali et al; Proc. Natl. Acad. Sci. USA, 79: 4927 (1982), Herpesviruses (E.P.A. 0473210A2), and Retroviruses (Valerio, D. et al; in Baum, S.J., Dicke,
- K.A., Lotzova, E. and Pluznik, D.H. (Eds.), Experimental Haematology today 1988. Springer Verlag, New York: pp. 92-99 (1989)).

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- Viruses known and used in the art as very suitable vector viruses specifically in poultry are Fowlpox virus, Marek's serotype 3 virus, Herpes virus of Turkey, Semliki Forest virus and Newcastle Disease virus.
- Live Recombinant Carriers are also known in the art as "live vectors", or shortly "vectors". Vaccines based upon a Live Recombinant Carrier are therefore also known in the art as vector vaccines.
- 35 The technique of *in vivo* homologous recombination, well-known in the art, can be used to introduce a recombinant nucleic acid into the genome of a bacterium, parasite or virus of

choice, capable of inducing expression of the inserted nucleic acid according to the invention in the host animal.

Finally another form of this embodiment of the invention relates to a host cell comprising a nucleic acid encoding a protein according to the invention, a DNA fragment comprising such a nucleic acid or a recombinant DNA molecule comprising such a nucleic acid under the control of a functionally linked promoter. This form also relates to a host cell containing a live recombinant carrier comprising a nucleic acid molecule encoding an *Ornithobacterium* rhinotracheale protein or an immunogenic fragment thereof according to the invention.

A host cell may be a cell of bacterial origin, e.g. Escherichia coli, Bacillus subtilis and Lactobacillus species, in combination with bacteria-based plasmids as pBR322, or bacterial expression vectors as pGEX, or with bacteriophages. The host cell may also be of eukaryotic origin, e.g. yeast-cells in combination with yeast-specific vector molecules, or higher eukaryotic cells like insect cells (Luckow et al; Bio-technology 6: 47-55 (1988)) in combination with vectors or recombinant baculoviruses, plant cells in combination with

combination with vectors or recombinant baculoviruses, plant cells in combination with e.g. Ti-plasmid based vectors or plant viral vectors (Barton, K.A. et al; Cell 32: 1033 (1983), mammalian cells like Hela cells, Chinese Hamster Ovary cells (CHO) or Crandell Feline Kidney-cells, also with appropriate vectors or recombinant viruses.

Another embodiment of the invention relates to an *Ornithobacterium rhinotracheale* protein and to immunogenic fragments thereof according to the invention.

The concept of immunogenic fragments will be defined below.

One form of this embodiment relates to a 59.8 kD Ornithobacterium rhinotracheale protein and to immunogenic fragments thereof, having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 2.

In a preferred form, the embodiment relates to such Ornithobacterium rhinotracheale proteins and immunogenic fragments thereof, that have a sequence homology of at least 85 %, preferably 90 %, more preferably 95 % homology to the amino acid sequence as depicted in SEQ ID NO: 2.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

The level of protein homology can be determined with the computer program "BLAST 2 SEQUENCES" by selecting sub-program: "BLASTP", that can be found at www.ncbi.nlm.nih.gov/blast/bl2seq/bl2.html.

A reference for this program is Tatiana A. Tatusova, Thomas L. Madden FEMS Microbiol. Letters 174: 247-250 (1999). Matrix used: "blosum62". Parameters used are the default parameters:

Open gap: 11. Extension gap: 1. Gap x_dropoff: 50.

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Another form of this embodiment relates to a 58.2 kD Ornithobacterium rhinotracheale protein and to immunogenic fragments thereof, having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 4.

In a preferred form, the embodiment relates to such *Ornithobacterium rhinotracheale* proteins and immunogenic fragments thereof, that have a sequence homology of at least 85 %, preferably 90 %, more preferably 95 % homology to the amino acid sequence as depicted in SEQ ID NO: 4.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

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Still another form of this embodiment relates to a 46.0 kD Ornithobacterium rhinotracheale protein and to immunogenic fragments thereof, having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 6.

In a preferred form, the embodiment relates to such *Ornithobacterium rhinotracheale* proteins and immunogenic fragments thereof, that have a sequence homology of at least 85 %, preferably 90 %, more preferably 95 % homology to the amino acid sequence as depicted in SEQ ID NO: 6.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

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Again another form of this embodiment relates to a 37.2 kD Ornithobacterium rhinotracheale protein and to immunogenic fragments thereof, having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 8.

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In a preferred form, the embodiment relates to such *Ornithobacterium rhinotracheale* proteins and immunogenic fragments thereof, that have a sequence homology of at least 85 %, preferably 90 %, more preferably 95 % homology to the amino acid sequence as depicted in SEQ ID NO: 8.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

Still another form of this embodiment relates to a 45.6 kD Ornithobacterium rhinotracheale protein and to immunogenic fragments thereof, having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 10.

In a preferred form, the embodiment relates to such *Ornithobacterium rhinotracheale* proteins and immunogenic fragments thereof, that have a sequence homology of at least 85 %, preferably 90 %, more preferably 95 % homology to the amino acid sequence as depicted in SEQ ID NO: 10.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

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One other form of this embodiment relates to a 42.2 kD Ornithobacterium rhinotracheale protein and to immunogenic fragments thereof, having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 12.

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In a preferred form, the embodiment relates to such *Ornithobacterium rhinotracheale* proteins and immunogenic fragments thereof, that have a sequence homology of at least 85 %, preferably 90 %, more preferably 95 % homology to the amino acid sequence as depicted in SEQ ID NO: 12.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

And again another form of this embodiment relates to a 34.0 kD Ornithobacterium rhinotracheale protein and to immunogenic fragments thereof, having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 14.

In a preferred form, the embodiment relates to such *Ornithobacterium rhinotracheale* proteins and immunogenic fragments thereof, that have a sequence homology of at least 85 %, preferably 90 %, more preferably 95 % homology to the amino acid sequence as depicted in SEQ ID NO: 14.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

Finally another form of this embodiment relates to a 32.9 kD Ornithobacterium rhinotracheale protein and to immunogenic fragments thereof, having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO:

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In a preferred form, the embodiment relates to such *Ornithobacterium rhinotracheale* proteins and immunogenic fragments thereof, that have a sequence homology of at least 85 %, preferably 90 %, more preferably 95 % homology to the amino acid sequence as depicted in SEQ ID NO: 16.

Even more preferred is a homology level of 98 %, 99 % or even 100 %.

Another form of this embodiment relates to such Ornithobacterium rhinotracheale proteins and immunogenic fragments of said proteins according to the invention, wherein the proteins and immunogenic fragments thereof are encoded by a nucleic acid according to the invention.

It will be understood that, for the particular proteins embraced herein, natural variations can exist between individual Ornithobacterium rhinotracheale strains. These variations may be demonstrated by (an) amino acid difference(s) in the overall sequence or by deletions, substitutions, insertions, inversions or additions of (an) amino acid(s) in said sequence. Amino acid substitutions which do not essentially alter biological and immunological activities, have been described, e.g. by Neurath et al in "The Proteins" Academic Press New York (1979). Amino acid replacements between related amino acids or replacements which have occurred frequently in evolution are, inter alia, Ser/Ala, Ser/Gly, Asp/Gly, Asp/Asn, Ile/Val (see Dayhof, M.D., Atlas of protein sequence and structure, Nat. Biomed. Res. Found., Washington D.C., 1978, vol. 5, suppl. 3). Other amino acid substitutions include Asp/Glu, Thr/Ser, Ala/Gly, Ala/Thr, Ser/Asn, Ala/Val, Thr/Phe, Ala/Pro, Lys/Arg, Leu/Ile, Leu/Val and Ala/Glu. Based on this information, Lipman and Pearson developed a method for rapid and sensitive protein comparison (Science, 227, 1435-1441, 1985) and determining the functional similarity between homologous proteins. Such amino acid substitutions of the exemplary embodiments of this invention, as well as variations having deletions and/or insertions are within the scope of the invention as long as the resulting proteins retain their immune reactivity.

This explains why Ornithobacterium rhinotracheale proteins according to the invention, when isolated from different field isolates, may have homology levels as low as about 80%, while still representing the same protein with the same immunological characteristics.

Those variations in the amino acid sequence of a certain protein according to the invention that still provide a protein capable of inducing an immune response against infection with

Ornithobacterium rhinotracheale or at least against the clinical manifestations of the infection are considered as "not essentially influencing the immunogenicity".

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When a protein is used for e.g. vaccination purposes or for raising antibodies, it is however not necessary to use the whole protein. It is also possible to use a fragment of that protein that is capable, as such or coupled to a carrier such as e.g. KLH, of inducing an immune response against that protein, a so-called immunogenic fragment. An "immunogenic fragment" is understood to be a fragment of the full-length protein that still has retained its capability to induce an immune response in a vertebrate host, e.g. comprises a B- or T-cell epitope. Shortly, an immunogenic fragment is a fragment that is capable of inducing an antigenic response against an Ornithobacterium rhinotracheale protein according to the invention. At this moment, a variety of techniques is available to easily identify DNA fragments encoding antigenic fragments (determinants). The method described by Geysen et al (Patent Application WO 84/03564, Patent Application WO 86/06487, US Patent NR. 4,833,092, Proc. Natl Acad. Sci. 81: 3998-4002 (1984), J. Imm. Meth. 102, 259-274 (1987), the so-called PEPSCAN method is an easy to perform, quick and well-established method for the detection of epitopes; the immunologically important regions of the protein. The method is used worldwide and as such well-known to man skilled in the art. This (empirical) method is especially suitable for the detection of B-cell epitopes. Also, given the sequence of the gene encoding any protein, computer algorithms are able to designate specific protein fragments as the immunologically important epitopes on the basis of their sequential and/or structural agreement with epitopes that are now known. The determination of these regions is based on a combination of the hydrophilicity criteria according to Hopp and Woods (Proc. Natl. Acad. Sci. 78: 38248-3828 (1981)), and the secondary structure aspects according to Chou and Fasman (Advances in Enzymology 47: 45-148 (1987) and US Patent 4,554,101). T-cell epitopes can likewise be predicted from the sequence by computer with the aid of Berzofsky's amphiphilicity criterion (Science 235, 1059-1062 (1987) and US Patent application NTIS US 07/005,885). A condensed overview is found in: Shan Lu on common principles: Tibtech 9: 238-242 (1991), Good et al on Malaria epitopes; Science 235: 1059-1062 (1987), Lu for a review; Vaccine 10: 3-7 (1992), Berzofsky for HIV-epitopes; The FASEB Journal 5:2412-2418 (1991). An immunogenic fragment usually has a minimal length of 8 amino acids, preferably more then 8, such as 9, 10, 12, 15 or even 20 amino acids. The nucleic acids encoding such a fragment therefore have a length of at least 24, but preferably 27, 30, 36, 45 or even 60 nucleic acids.

Therefore, one form of still another embodiment of the invention relates to vaccines for combating *Ornithobacterium rhinotracheale* infection, that comprise an *Ornithobacterium rhinotracheale* protein or immunogenic fragments thereof, according to the invention as described above together with a pharmaceutically acceptable carrier.

Still another embodiment of the present invention relates to an *Ornithobacterium* rhinotracheale protein according to the invention or immunogenic fragments thereof for use in a vaccine.

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Still another embodiment of the present invention relates to the use of a nucleic acid, a DNA fragment, a recombinant DNA molecule, a live recombinant carrier, a host cell or a protein or an immunogenic fragment thereof according to the invention for the manufacturing of a vaccine for combating *Ornithobacterium rhinotracheale* infection.

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One way of making a vaccine according to the invention is by growing the bacteria, followed by biochemical purification of an *Ornithobacterium rhinotracheale* protein or an immunogenic fragment thereof, from the bacterium. This is however a very time-consuming way of making the vaccine.

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It is therefore much more convenient to use the expression products of the gene encoding an Ornithobacterium rhinotracheale protein or immunogenic fragments thereof in vaccines. This is possible for the first time now because the nucleic acids encoding the Ornithobacterium rhinotracheale proteins are provided in the present invention.

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Vaccines based upon the expression products of these genes can easily be made by admixing the protein according to the invention or immunogenic fragments thereof according to the invention with a pharmaceutically acceptable carrier as described below.

Alternatively, a vaccine according to the invention can comprise live recombinant carriers as described above, capable of expressing the protein according to the invention or immunogenic fragments thereof. Such vaccines, e.g. based upon a Salmonella carrier or a viral carrier e.g. a Herpesvirus vector have the advantage over subunit vaccines that they better mimic the natural way of infection of Ornithobacterium rhinotracheale. Moreover, their self-propagation is an advantage since only low amounts of the recombinant carrier are necessary

for immunization.

Vaccines can also be based upon host cells as described above, that comprise the protein or immunogenic fragments thereof according to the invention.

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All vaccines described above contribute to active vaccination, i.e. they trigger the host's defense system.

Alternatively, antibodies can be raised in e.g. rabbits or can be obtained from antibody-producing cell lines as described below. Such antibodies can then be administered to the chicken. This method of vaccination, passive vaccination, is the vaccination of choice when an animal is already infected, and there is no time to allow the natural immune response to be triggered. It is also the preferred method for vaccinating animals that are prone to sudden high infection pressure. The administered antibodies against the protein according to the invention or immunogenic fragments thereof can in these cases bind directly to *Ornithobacterium rhinotracheale*. This has the advantage that it decreases or stops *Ornithobacterium rhinotracheale* multiplication.

Therefore, one other form of this embodiment of the invention relates to a vaccine for combating *Ornithobacterium rhinotracheale* infection that comprises antibodies against a *Ornithobacterium rhinotracheale* protein according to the invention or an immunogenic fragment of that protein, and a pharmaceutically acceptable carrier.

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- Still another embodiment of this invention relates to antibodies against a *Ornithobacterium* rhinotracheale protein according to the invention or an immunogenic fragment of that protein.
- Methods for large-scale production of antibodies according to the invention are also known in 20 the art. Such methods rely on the cloning of (fragments of) the genetic information encoding the protein according to the invention in a filamentous phage for phage display. Such techniques are described i.a. at the "Antibody Engineering Page" under "filamentous phage display" at http://aximtl.imt.uni-marburg.de/~rek/aepphage.html., and in review papers by Cortese, R. et al., (1994) in Trends Biotechn. 12: 262-267., by Clackson, T. & Wells, J.A. (1994) in Trends Biotechn. 12: 173-183, by Marks, J.D. et al., (1992) in J. Biol. Chem. 267: 25 16007-16010, by Winter, G. et al., (1994) in Annu. Rev. Immunol. 12: 433-455, and by Little, M. et al., (1994) Biotechn. Adv. 12: 539-555. The phages are subsequently used to screen camelid expression libraries expressing camelid heavy chain antibodies. (Muyldermans, S. and Lauwereys, M., Journ. Molec. Recogn. 12: 131-140 (1999) and Ghahroudi, M.A. et al., 30 FEBS Letters 414: 512-526 (1997)). Cells from the library that express the desired antibodies can be replicated and subsequently be used for large scale expression of antibodies.

Still another embodiment relates to a method for the preparation of a vaccine according to the invention that comprises the admixing of antibodies according to the invention and a pharmaceutically acceptable carrier.

An alternative and efficient way of vaccination is direct vaccination with DNA encoding the relevant antigen. Direct vaccination with DNA encoding proteins has been successful for many different proteins. (As reviewed in e.g. Donnelly et al., The Immunologist 2: 20-26 (1993)). This way of vaccination is also attractive for the vaccination of chickens against *Ornithobacterium rhinotracheale* infection.

Therefore, still other forms of this embodiment of the invention relate to vaccines comprising nucleic acids encoding a protein according to the invention or immunogenic fragments thereof, comprising DNA fragments that comprise such nucleic acids or comprising recombinant DNA molecules according to the invention, and a pharmaceutically acceptable carrier.

Examples of DNA plasmids that are suitable for use in a DNA vaccine according to the invention are conventional cloning or expression plasmids for bacterial, eukaryotic and yeast host cells, many of said plasmids being commercially available. Well-known examples of such plasmids are pBR322 and pcDNA3 (Invitrogen). The DNA fragments or recombinant DNA molecules according to the invention should be able to induce protein expression of the nucleotide sequences. The DNA fragments or recombinant DNA molecules may comprise one or more nucleotide sequences according to the invention. In addition, the DNA fragments or recombinant DNA molecules may comprise other nucleotide sequences such as the immune-stimulating oligonucleotides having unmethylated CpG di-nucleotides, or nucleotide sequences that code for other antigenic proteins or adjuvating cytokines.

The nucleotide sequence according to the present invention or the DNA plasmid comprising a nucleotide sequence according to the present invention, preferably operably linked to a transcriptional regulatory sequence, to be used in the vaccine according to the invention can be naked or can be packaged in a delivery system. Suitable delivery systems are lipid vesicles, iscoms, dendromers, niosomes, polysaccharide matrices and the like, (see further below) all well-known in the art. Also very suitable as delivery system are attenuated live bacteria such as Salmonella species, and attenuated live viruses such as Herpesvirus vectors, as mentioned above.

DNA vaccines can e.g. easily be administered through intradermal application such as by using a needle-less injector. This way of administration delivers the DNA directly into the cells of the animal to be vaccinated. Amounts of DNA in the range between 10 pg and 1000 μ g provide good results. Preferably, amounts in the microgram range between 1 and 100 μ g are used.

In a further embodiment, the vaccine according to the present invention comprises one or more additional antigens derived from a virus or micro-organism pathogenic to poultry, an antibody against such an antigen or genetic information encoding said antigen.

- Of course, such antigens can be e.g. other *Ornithobacterium rhinotracheale* antigens. It is beneficial to combine, in one vaccine, two or more of the proteins or immunogenic fragments thereof according to the invention, antibodies against such proteins or immunogenic fragments thereof, or genetic information encoding such proteins or immunogenic fragments thereof.
- Next to this, it is beneficial to include in a vaccine according to the invention, antigens derived from another micro-organism or a virus pathogenic to poultry, an antibody against such an antigen or genetic information encoding said antigen.
- Preferably, the virus or micro-organism is selected from the group consisting of Fowlpox virus, Infectious Bronchitis virus, Infectious Bursal Disease (Gumboro), Marek's Disease Virus, Chicken Anaemia agent, Avian Reovirus, Mycoplasma gallisepticum, Turkey Rhinotracheitis virus, Haemophilus paragallinarum (Coryza), Chicken Poxvirus, Avian Encephalomyelitisvirus, Duck Plague virus, Newcastle Disease virus, Egg Drop syndrome virus, Infectious Laryngotracheitis virus, Herpes Virus of Turkeys, Eimeria species,

 Ornithobacterium rhinotracheale, Pasteurella multocida, Mycoplasma smoviga, Salmore Virus
- Ornithobacterium rhinotracheale, Pasteurella multocida, Mycoplasma synoviae, Salmonella species and E. coli.
- Vaccines based upon the *Ornithobacterium rhinotracheale* proteins according to the invention are also very suitable as marker vaccines. A marker vaccine is a vaccine that allows to

 25 discriminate between vaccinated and field-infected chickens e.g. on the basis of a characteristic antibody panel, different from the antibody panel induced by wild type infection. A different antibody panel is induced e.g. when an immunogenic protein present on a wild type bacterium is not present in a vaccine: the host will then not make antibodies against that protein after vaccination. Thus, a vaccine based upon an *Ornithobacterium*30 *rhinotracheale* protein according to the invention would only induce antibodies against that protein, whereas a vaccine based upon a live wild-type, live attenuated or inactivated whole *Ornithobacterium rhinotracheale* would induce antibodies against all or most of the bacterial proteins.
- A simple ELISA test, having wells comprising one protein according to the invention and wells comprising another protein according to the invention suffices to test serum from chickens and to tell if the chickens are either vaccinated with a subunit vaccine according to the invention or suffered from *Ornithobacterium rhinotracheale* field infection; chickens

vaccinated with a vaccine comprising one protein according to the invention would not have antibodies against another protein according to the invention. Chickens that have encountered a field infection with *Ornithobacterium rhinotracheale* would however have antibodies against all immunogenic *Ornithobacterium rhinotracheale* proteins and thus also against another protein according to the invention.

All vaccines according to the present invention comprise a pharmaceutically acceptable carrier. A pharmaceutically acceptable carrier can be e.g. sterile water or a sterile physiological salt solution. In a more complex form the carrier can e.g. be a buffer.

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Methods for the preparation of a vaccine comprise the admixing of a protein or an immunogenic fragment thereof, according to the invention and/or antibodies against that protein or an immunogenic fragment thereof, and/or a nucleic acid and/or a DNA fragment, a recombinant DNA molecule, a live recombinant carrier or host cell according to the invention, and a pharmaceutically acceptable carrier.

Vaccines according to the present invention may in a preferred presentation also contain an immunostimulatory substance, a so-called adjuvant. Adjuvants in general comprise substances that boost the immune response of the host in a non-specific manner. A number of different adjuvants are known in the art. Examples of adjuvants frequently used in chicken vaccines are muramyldipeptides, lipopolysaccharides, several glucans and glycans and Carbopol^(R) (a homopolymer).

The vaccine may also comprise a so-called "vehicle". A vehicle is a compound to which the protein adheres, without being covalently bound to it. Such vehicles are i.a. bio-microcapsules, micro-alginates, liposomes and macrosols, all known in the art.

A special form of such a vehicle, in which the antigen is partially embedded in the vehicle, is the so-called ISCOM (EP 109.942, EP 180.564, EP 242.380)

In addition, the vaccine may comprise one or more suitable surface-active compounds or emulsifiers, e.g. Span or Tween.

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Often, the vaccine is mixed with stabilisers, e.g. to protect degradation-prone proteins from being degraded, to enhance the shelf-life of the vaccine, or to improve freeze-drying efficiency. Useful stabilisers are i.a. SPGA (Bovarnik et al; J. Bacteriology 59: 509 (1950)), carbohydrates e.g. sorbitol, mannitol, trehalose, starch, sucrose, dextran or glucose, proteins such as albumin or casein or degradation products thereof, and buffers, such as alkali metal phosphates.

In addition, the vaccine may be suspended in a physiologically acceptable diluent.

It goes without saying, that other ways of adjuvating, adding vehicle compounds or diluents, emulsifying or stabilising a protein are also embodied in the present invention.

Vaccines according to the invention that are based upon the protein according to the invention or immunogenic fragments thereof can very suitably be administered in amounts ranging between 1 and 100 micrograms of protein per animal, although smaller doses can in principle be used. A dose exceeding 100 micrograms will, although immunologically very suitable, be less attractive for commercial reasons.

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Vaccines based upon live attenuated recombinant carriers, such as the LRC-viruses and bacteria described above can be administered in much lower doses, because they multiply themselves during the infection. Therefore, very suitable amounts would range between 10³ and 10⁹ CFU/PFU for respectively bacteria/viruses.

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Vaccines according to the invention can be administered e.g. intradermally, subcutaneously, intramuscularly, intraperitoneally, intravenously, or at mucosal surfaces such as orally or intranasally.

Live recombinant carrier vaccines or vector vaccines can most efficiently be administered by spraying, by aerosol or by drinking water administration.

Examples.

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Example 1: Library construction, sera and screening.

For the construction of an expression library of *Ornithobacterium rhinotracheale* serotype G strain O-95029 nr.16279, genomic DNA was isolated from cells grown in Todd Hewitt broth (THB) for 24 hours at 37°C on a 100 rpm shaker, according to the method described in Maniatis/Sambrook (Sambrook, J. *et al.* Molecular cloning: a laboratory manual. ISBN 0-87969-309-6). DNA fragments of 1 – 4 kb were obtained by restriction enzyme digestion and ligated into λTriplEx vector arms (Clontech, Palo Alto, CA, USA). Subsequent packaging was performed using the Stratagene (La Jolla, CA, USA) *in vitro* packaging extract. *Escherichia coli* XL1 Blue cells, grown in Luria Bertani (LB) broth supplemented with 10 mM MgSO₄ and 0.2% maltose, were used for transfection. The complexity of the constructed expression library was tested 6.9 and it contained 97% recombinants.

The Ornithobacterium rhinotracheale serotype G expression library was screened with polyclonal antisera directed against whole live organisms of several Ornithobacterium rhinotracheale serotypes. Sera were collected from broiler chickens that were vaccinated by aerosol spraying with live Ornithobacterium rhinotracheale bacteria of serotype B (strain GGD 1261), serotype G (strain O-95029 nr.16279) or serotype M (strain TOP 98036 4500) at two weeks of age. Three weeks later the chickens were intravenously challenged with Ornithobacterium rhinotracheale serotype A (strain B3263/91). Sera were collected one week after challenge. All vaccinated birds showed reduced pathology (ranging from 10% to 60%) in comparison to unvaccinated control birds. Before use in expression library screening, the antisera were adsorbed with Escherichia coli XL1 Blue cell lysate as described in Maniatis/Sambrook (Sambrook, J. et al. Molecular cloning: a laboratory manual. ISBN 0-87969-309-6) in order to reduce a-specific background signal.

The expression library was screened by plaque lift using an initial screening of approximately 20.000 plaques. The procedure was done as described in the manufacturers handbook (Clontech, Palo Alto, CA, USA). All library screenings were done under native conditions.

In short, phage-infected Escherichia coli XL1 Blue cells were plated in LB top agar onto LB agar plates both supplemented with 10 mM MgSO₄. The plates were then incubated at 42°C for 4 hours. A nitrocellulose filter disc (Schleicher and Schuell, Dassel, Germany), previously soaked in 10 mM IPTG, was placed on each plate in order to induce expression of the proteins encoded by the cloned Ornithobacterium rhinotracheale inserts. After 4 hours incubation at 37°C all filters were removed from the plates. After washing and blocking, filters were

incubated with chicken antiserum (pooled from 10 animals, 1:250 dilution). The antiserum used in the first screening was obtained from chickens live vaccinated with *Ornithobacterium rhinotracheale* serotype G followed by a challenge with *Ornithobacterium rhinotracheale* serotype A. As secondary antibody rabbit anti-chicken IgG peroxidase (Nordic, Tilburg, The Netherlands) was use at 1:1000 dilution. As substrate solution Vector SG (Vector, Burlingame, CA, USA) was used.

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From the initial screening of 20.000 plaques, 200 reactive plaques were located on the agar plates and isolated. A plaque lift and screen as described above was repeated twice resulting in 175 single, pure reactive plaques. The pure clones were then spotted *in duplo* onto an

- E.coli XL1 Blue top agar lawn to give confluent plaques of approximately 5 mm diameter. Again a plaque lift was performed and the filters were incubated with the antisera obtained from birds live vaccinated with Ornithobacterium rhinotracheale serotype B or serotype M prior to Ornithobacterium rhinotracheale serotype A challenge. Out of 175 reactive plaques, 30 plaques were selected to be cross-reactive with sera from birds live vaccinated with
- Ornithobacterium rhinotracheale serotype B, serotype G, or serotype M, and challenged with Ornithobacterium rhinotracheale serotype A.

Example 2: Identification of open reading frames (ORFs) encoding antigenic proteins and expression in *Escherichia coli*.

- 20 The DNA inserts of the 30 selected plaques were analysed in order to identify the open reading frames encoding the antigenic proteins. Oligonucleotide primers designed for the λTriplEx vector arms were used for both PCR amplification and sequencing. PCR was performed in a final reaction volume of 50 µl containing 50 µM dNTP's (Promega, WI, USA), 10 pmol of both primers, 20 U/ml Supertaq plus polymerase and 10X Supertaq buffer 25 (both HT Biotechnology Ltd, Cambridge, UK) in water. Phage DNA was added by picking a freshly plated plaque using a tooth pick, and transferring this DNA from tooth pick to reaction mix. The following conditions were used: denaturation at 94°C for 3 min, followed by 30 cycles of denaturation at 94°C for 1 min, annealing at 50°C for 2 min and elongation at 68°C for 2 min 30 sec, followed by a final extension at 68°C for 10 min. To determine the 30 nucleotide sequence of the amplified DNA inserts a sequence reaction was done (94°C 10 sec; 50°C 5 sec; 60°C 2 min for 25 cycles) using Big dye Terminator Ready reaction mix (Qiagen Inc., CA, USA), 50 ng template DNA (PCR product) and 2.4 pmol primer in a 20µl reaction volume.
- After sequence analysis the 30 clones appeared to represent 8 different genes. Since most open reading frames where a fusion with the lacZ gene of the λTriplEx vector, the 5'end of

the gene was missing. For that reason a sequence reaction was performed using internal primers and chromosomal DNA of *Ornithobacterium rhinotracheale* serotype G as a template to sequence the missing 5'gap.

Oligonucleotide primers were designed to amplify the full length open reading frames 5 encoding the 8 cross-reactive antigens (Or01, Or02, Or03, Or04, Or11, Or77, Or98A and Or98B) from genomic DNA of Ornithobacterium rhinotracheale serotype G strain O-95029 nr.16279 (see table 1). The 5'oligonucleotide primers contain a restriction site (underlined) preceding the ATG initiation codon (bold) followed by sequences derived from the gene of interest (italic). The 3'oligonucleotides contain coding sequences (italic) followed by a 10 restriction site (underlined). The PCR products were cloned in the expression vector of interest. Ligation products were transformed to E.coli BL21 (DE3) codon RIL pLysS host cells (Novagen, Madison, WI, USA) for protein expression. By using the pET plasmid vector (pET22b) and a T7 RNA polymerase expression system (Novagen, Madison, WI, USA), the recombinant proteins were expressed in E.coli, with an E.coli pelB leader peptide fused at the 15 amino terminal portion (Ornithobacterium rhinotracheale leader peptides of proteins Or02, Or03, Or11, and Or77 were replaced) and 6 histidine residues at the carboxy terminal portion of the protein. E.coli strain BL21 (DE3) codon RIL pLysS (Novagen, Madison, WI, USA) was used for high level expression during IPTG-induction as described in the pET system manual (Novagen, Madison, WI, USA). 20

Example 3: Purification of antigens, vaccine formulations and serological analysis.

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Recombinant antigens expressed in *E.coli* were isolated from supernatant (Or77), purified by metal affinity chromatography using talon resin (Clontech Inc., Palo Alto, CA, USA) as described by the manufacturer (Or03, Or04, Or98A and Or98B), or by repeated freeze-thawing, sonification, and centrifugation cycli (Or01, Or02 and Or11). Polyacrylamide gel electrophoresis (PAGE) followed by Coomassie brilliant blue staining was used to assess the purity of the recombinant proteins. Protein concentrations were estimated using bovine serum albumin as the standard.

All purified recombinant proteins (Or01, Or02, Or03, Or04, Or11, Or77, Or98A and Or98B) were formulated individually in a water in oil emulsion. Furthermore, five different subunit vaccines (A, B, C, D and E) were formulated, containing different compositions of the 8 recombinant antigens (table 2). Coomassie staining of the 5 combination vaccines showed clearly identifiable protein bands corresponding to recombinant proteins Or01, Or02 and

Or77. As the molecular weights of Or03, Or04 and Or11, and the molecular weights of Or98A and Or98B are approximately the same, individual protein bands could not be distinguished (figure 1). All proteins are present in approximately equal concentrations of 50 mg/antigen/l (25 µg/dose). Therefore, the total antigenic load of vaccine A to D is 200 mg/l. The antigen concentration of vaccine E is 400 mg/l. The protein background is rest material from *E.coli* strain used to express the recombinant *Ornithobacterium rhinotracheale* antigens.

The ability of the different subunit vaccines to stimulate the humoral immune response to produce protein-specific antibodies was studied by subcutaneous injection of 2-weeks-old SPF-broiler chickens with 0.5 ml vaccine. Four weeks after vaccination serum-samples were collected and tested for the presence of antibodies reactive against the recombinant proteins. Semi-dry Western blotting was performed according to Towbin, H., Staehlin, T., and Gordon, J. (1979) Proc. Nat. Acad. Sci. 76:43-50. The protein phase of the vaccines was blotted and incubated with pooled serum (1:100 dilution) from vaccinated and unvaccinated birds. Sera obtained from birds vaccinated with each of the 8 individual vaccines Or01 to Or98B showed protein-specific reactivity (figure 2). Figure 3 shows the reactivity of antisera obtained from birds vaccinated with subunit vaccine A to E (see table 2 and figure 1), directed against the same vaccines on Western blot. For example: blot A is loaded with vaccine A, B, C, D, and E (corresponding with lanes A to E). The serum used for primary antibody binding is obtained from birds vaccinated with vaccine A (corresponds with blot-number). For this reason, a-Or01, α -Or02, α -Or03 and α -Or04 antibodies are present in this serum. On blot A, these four proteins are stained in lane A, D, and E, which are the lanes that were loaded with the three vaccines that contain these antigens (A, D, and E). Blot B is loaded as blot A and the serum used is obtained from birds vaccinated with vaccine B. α -Or77, α -Or11, α -Or03, and α -Or04 antibodies stain the corresponding antigens on blot B in lane B, C, and E. The other antigens that were not present in vaccine B could not be detected on this blot. On blot E, all proteins are stained because vaccine E contains all eight Ornithobacterium rhinotracheale antigens. The serum used on Westernblot F is obtained from unvaccinated birds that served as a negative control. No recombinant Ornithobacterium rhinotracheale antigens could be detected using this serum.

Example 4: Protection studies.

To assess the cross-protective capacity of the antibody response induced by different subunit vaccines (combi vaccines A, B, C, D, E, and individual vaccine Or77), an animal experiment was performed. SPF-broilers were vaccinated at 2 weeks of age as described before. At 5

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weeks of age birds were primed with ND LaSota (dose: 1*106 E.I.D.per bird) by aerosol spraying. At 6 weeks of age, birds were challenged with Ornithobacterium rhinotracheale serotype A strain B3263/91 (heterologous challenge). The challenge was done by aerosol spraying of a fresh bacterial culture containing 8.5*10⁸ colony forming units (CFU) per ml THB. During aerosol challenge the bacterial culture was administered as a fine spray to the 5 birds in an isolator of approximately 1.5m³, using a commercial paint sprayer. The developed mist in the isolators was maintained for at least 10 min with the air circulation closed. Challenge control groups and ND priming groups were included in the test. One week after challenge, at 7 weeks of age, birds were sacrificed and organ lesions were macroscopically scored using an Ornithobacterium rhinotracheale scoring system for respiratory disease as 10 follows: for thoracic air sacs, 0= no abnormalities, 1= one air sac seriously affected by fibrinous airsacculitis or limited pin-head sized foci of fibrinous exudates in both air sacs, 2= both air sacs seriously affected by fibrinous airsacculitis; for abdominal air sacs, 0= no abnormalities, 1= pin-head sized foci of fibrinous exudates or slight diffuse fibrinous airsacculitis, 2= severe fibrinous airsacculitis. The airsacculitis score is given as the sum of 15 both scores. For lungs, 0= no abnormalities, 1= unilateral pneumonia, 2= bilateral pneumonia. The average group scores are given as a percentage of the maximum possible score. Statistical analysis was performed using Kruskal-Wallis non-parametric one-way ANOVA. Figure 4 shows the cross-protective capacity of the 5 different subunit vaccines A to E. The challenge control group was not vaccinated but primed and challenged and showed the 20 highest score. Birds vaccinated with vaccine E (containing all 8 antigens) showed almost complete protection comparable to the results of the group that did not receive vaccination and challenge but was primed with Newcastle Disease virus. A somewhat lesser, but still significant cross-protection (P<0.05) could be observed in birds vaccinated with vaccine A, B and C. Combination vaccine D showed cross-protection of less significance (p=0.19). 25 Untreated birds showed no organ lesions. As can be seen from figure 5, the Or77 (= serotype G strain)-vaccinated and serotype A challenged animals also show a significant (p<0.05)) reduction in respiratory lesion scores compared to the unvaccinated control group.

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Legend to the figures:

Figure 1: Coomassie staining of the 5 combination vaccines (A to E). Each vaccine containing a different composition of the 8 purified recombinant proteins. Subunit vaccine A corresponds with lane A, subunit vaccine B corresponds with lane B, subunit vaccine C corresponds with lane C, subunit vaccine D corresponds with lane D, subunit vaccine E corresponds with lane E. Recombinant proteins with approximately equal molecular weights are indicated by a single arrow.

Figure 2: Reactivity of monovalent antisera, obtained from chickens vaccinated with the single recombinant subunit vaccines, against the same protein on Western blot. The reactive vaccine proteins are indicated with black arrows.

Figure 3: Reactivity of antisera, obtained from chickens vaccinated with subunit vaccines A to E on Western blot. Each blot contains the proteins of vaccine A, B, C, D, and E (corresponding to lanes A to E). The serum used for screening is obtained from birds vaccinated with vaccine A (blot A), vaccine B (blot B), vaccine C (blot C), vaccine D (blot D) or vaccine E (blot E). The serum used on Western blot F is obtained from unvaccinated birds. The reactive vaccine proteins are indicated with a black line.

Figure 4: Cross-protective capacity of subunit vaccines A to E, in comparison to challenge and NDV control groups, represented as the maximum possible respiratory organ lesion score.

Figure 5: Cross-protective capacity of subunit vaccine Or77, in comparison to challenge and NDV control groups, represented as the maximum possible respiratory organ lesion score.

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Gene	5'oligonucleotide	Restriction	3'oligonucleotide	Restriction
		site		site
Or01	5'-GCTGGCCATGGCTGAAATTATAAAAATGCC-3'	MscI	5'-CCGCTCGAGCACATAGACATTGG-3'	Xhol
Or02	5'-CAGT <u>CCATGG</u> CATGTAGCGATTTTGAT-3'	Ncol	5'-CCGCTCGAGGTGGTCTTTATAAAATG-3'	Xhol
Or03	5'-CAGTCCATGGCGATGATAATCAGTTCTTATG-3'	Ncol	5'-CCGCTCGAGAATAAATTCATCATTAAGC-3'	Xhol
Or04	5'-CGATGGCCATGAAAGATATTTGAAT-3'	Mscl	5'-CCGCTCGAGTTCTTCACTTGGTATTTTGA-3'	Xhol
Or11	5'-CGATGGCCATGGGGCACAAGGTGTAGC-3'	Msci	5'-GCGGCCGC TACGATAAACCTAGACCAAA-3'	Noti
Or77	5'-CATGCCATGGTCTGTAGCAGTGATTAC-3'	Ncol	5'-CCGCTCGAGGTTAATTGAAACTCTTAAGC-3'	Xhol
Or98A	5'-CAGTCCATGGTAAAAGACTTTTCAG-3'	Ncol	5'-CCGCTCGAGTGCTATTAATTCTAATCG-3'	Xhol
Or98B	5'-CAGTCCATGGAATTAGCGAAAAACGAC-3'	Ncol	5'-CCGCTCGAGTTTTAATTCATTTTTTCTG-3'	Xhol

Restriction site: underlined

ATG start codon: bold

Gene of interest: italic

Table 1: Oligonucleotide sets used for cloning selected Ornithobacterium rhinotracheale genes encoding cross-reactive antigens

	Antigen							
Vaccine	Or01	Or02	Or03	Or04	Or11	Or77	Or98A	Or98B
A	nu 4							0.50B
В				1 12 14				
С								Farget tage of the
D		3 12 14 3 7						
E					. Of the control of the			

: antigen is present in the vaccine

Table 2: Subunit vaccines (A to E) consisting of different protein subset combinations

EPO DG 11.02.2004 (106)

- 1) Nucleic acid encoding a 59.8 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein, said nucleic acid or said part thereof having at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 1.
- Nucleic acid or part thereof according to claim 1, characterized in that the sequence has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO:

 1.
- 3) Nucleic acid encoding a 58.2 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein, said nucleic acid or said part thereof having at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 3.
- 4) Nucleic acid or part thereof according to claim 3, characterized in that the sequence has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 3.
- 5) Nucleic acid encoding a 46.0 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein, said nucleic acid or said part thereof having at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 5.
- Nucleic acid or part thereof according to claim 5, characterized in that the sequence has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 5.
- 7) Nucleic acid encoding a 37.2 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein, said nucleic acid or said part thereof having at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 7.
- Nucleic acid or part thereof according to claim 7, characterized in that the sequence has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 7.
- 9) Nucleic acid encoding a 45.6 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein, said

- nucleic acid or said part thereof having at least 80 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 9.
- 10) Nucleic acid or part thereof according to claim 9, characterized in that the sequence has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 9.
- 11) Nucleic acid encoding a 42.2 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein, said nucleic acid or said part thereof having at least 80 % homology with the nucleic acid of he Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 11.
- 12) Nucleic acid or part thereof according to claim 11, characterized in that the sequence has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 11.
- 13) Nucleic acid encoding a 34.0 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein, said nucleic acid or said part thereof having at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 13.
- 14) Nucleic acid or part thereof according to claim 13, characterized in that the sequence has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 13.
- 15) Nucleic acid encoding a 32.9 kD Ornithobacterium rhinotracheale protein or a part of said nucleic acid that encodes an immunogenic fragment of said protein, said nucleic acid or said part thereof having at least 80 % homology with the nucleic acid of the Ornithobacterium rhinotracheale protein gene as depicted in SEQ ID NO: 15.
- 16) Nucleic acid or part thereof according to claim 15, characterized in that the sequence has at least 85 %, preferably 90 %, more preferably 95 % homology with the nucleic acid of the *Ornithobacterium rhinotracheale* protein gene as depicted in SEQ ID NO: 15.
- 17) DNA fragment comprising a nucleic acid according to claim 1-16.
- 18) Recombinant DNA molecule comprising a nucleic acid according to claims 1-16 or a DNA fragment according to claim 17, under the control of a functionally linked promoter.
- 19) Live recombinant carrier comprising a nucleic acid according to claims 1-16, a DNA fragment according to claim 17 or a recombinant DNA molecule according to claim 18.

- 20) Host cell comprising a nucleic acid according to claims 1-16, a DNA fragment according to claim 17, a recombinant DNA molecule according to claim 18 or a live recombinant carrier according to claim 19.
- 21) A 59.8 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 2.
- 22) A Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, according to claim 21, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 85 %, preferably 90 %, more preferably 95 % to the amino acid sequence as depicted in SEQ ID NO: 2.
- 23) A 59.8 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment thereof, characterized in that it is encoded by a nucleic acid according to claim 1 or 2.
- 24) A 58.2 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 80 % to the amino acid sequence as depicted in SEQ ID NO: 4.
- 25) A Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, according to claim 24, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 85 %, preferably 90 %, more preferably 95 % to the amino acid sequence as depicted in SEQ ID NO: 4.
- 26) A 58.2 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment thereof, characterized in that it is encoded by a nucleic acid according to claim 3 or 4.
- 27) A 46.0 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 6.
- 28) A Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, according to claim 27, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 85 %, preferably 90 %, more preferably 95 % to the amino acid sequence as depicted in SEQ ID NO: 6.
- 29) A 46.0 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment thereof, characterized in that it is encoded by a nucleic acid according to claim 5 or 6.
- 30) A 37.2 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 8.

- 31) A Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, according to claim 30, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 85 %, preferably 90 %, more preferably 95 % to the amino acid sequence as depicted in SEQ ID NO: 8.
- 32) A 37.2 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment thereof, characterized in that it is encoded by a nucleic acid according to claim 7 or 8.
- 33) A 45.6 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 10.
- 34) A Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, according to claim 33, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 85 %, preferably 90 %, more preferably 95 % to the amino acid sequence as depicted in SEQ ID NO: 10.
- 35) A 45.6 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment thereof, characterized in that it is encoded by a nucleic acid according to claim 9 or 10.
- 36) A 42.2 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 12.
- 37) A Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, according to claim 36, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 85 %, preferably 90 %, more preferably 95 % to the amino acid sequence as depicted in SEQ ID NO: 12.
- 38) A 42.2 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment thereof, characterized in that it is encoded by a nucleic acid according to claim 11 or 12.
- 39) A 34.0 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 14.
- 40) A Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, according to claim 39, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 85 %, preferably 90 %, more preferably 95 % to the amino acid sequence as depicted in SEQ ID NO: 14.

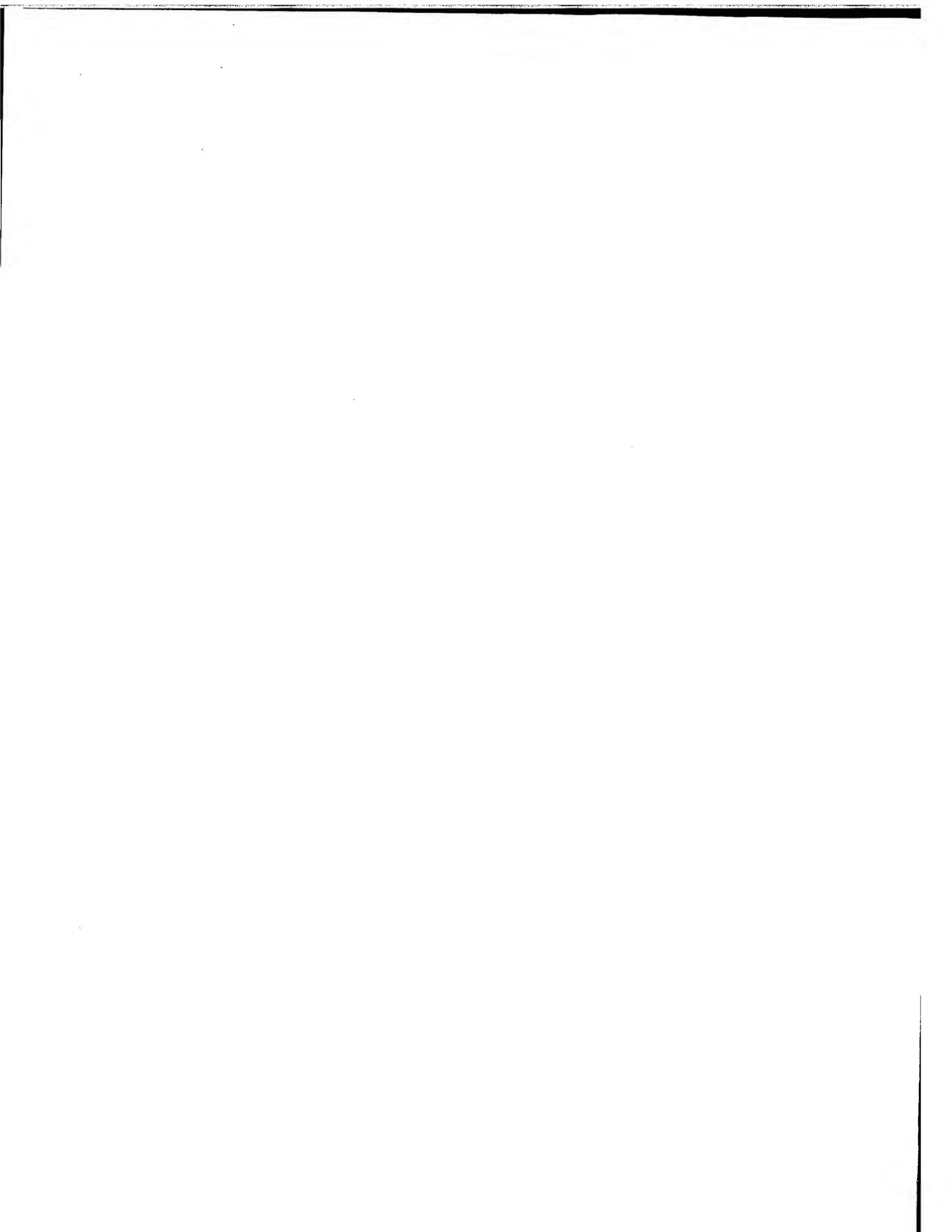
- 41) A 34.0 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment thereof, characterized in that it is encoded by a nucleic acid according to claim 13 or 14.
- 42) A 32.9 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 80 % with the amino acid sequence as depicted in SEQ ID NO: 16.
- 43) A Ornithobacterium rhinotracheale protein or an immunogenic fragment of said protein, according to claim 42, said protein or immunogenic fragment thereof having an amino acid sequence homology of at least 85 %, preferably 90 %, more preferably 95 % to the amino acid sequence as depicted in SEQ ID NO: 16.
- 44) A 32.9 kD Ornithobacterium rhinotracheale protein or an immunogenic fragment thereof, characterized in that it is encoded by a nucleic acid according to claim 15 or 16.
- 45) A nucleic acid according to claims 1-16, a DNA fragment according to claim 17, a recombinant DNA molecule according to claim 18, a live recombinant carrier according to claim 19, a host cell according to claim 20 or a protein according to claims 21-44 or an immunogenic fragment thereof, for use in a vaccine.
- 46) Use of a nucleic acid according to claims 1-16, a DNA fragment according to claim 17, a recombinant DNA molecule according to claim 18, a live recombinant carrier according to claim 19, a host cell according to claim 20 or a protein according to claims 21-44 or an immunogenic fragment thereof for the manufacturing of a vaccine for combating Ornithobacterium rhinotracheale infection.
- 47) Vaccine for combating *Ornithobacterium rhinotracheale* infection, characterized in that it comprises a nucleic acid according to claims 1-16, a DNA fragment according to claim 17, a recombinant DNA molecule according to claim 18, a live recombinant carrier according to claim 19, a host cell according to claim 20 or a protein according to claims 21-44 or an immunogenic fragment thereof, and a pharmaceutically acceptable carrier.
- 48) Vaccine for combating Ornithobacterium rhinotracheale infection, characterized in that it comprises antibodies against a protein according to claims 21-44 or an immunogenic fragment of said protein, and a pharmaceutically acceptable carrier.
- 49) Vaccine according to claim 47, characterized in that it comprises an adjuvant.
- 50) Vaccine according to claim 47-49, characterized in that it comprises an additional antigen derived from a virus or micro-organism pathogenic to poultry, an antibody against such an antigen or genetic information encoding said antigen.

- 51) Vaccine according to claim 50, characterized in that said virus or micro-organism pathogenic to chickens is selected from the group consisting of Fowlpox virus, Infectious Bronchitis virus, Infectious Bursal Disease (Gumboro), Marek's Disease Virus, Chicken Anaemia agent, Avian Reovirus, Mycoplasma gallisepticum, Turkey Rhinotracheitis virus, Haemophilus paragallinarum (Coryza), Chicken Poxvirus, Avian Encephalomyelitisvirus, Duck Plague virus, Newcastle Disease virus, Egg Drop syndrome virus, Infectious Laryngotracheitis virus, Herpes Virus of Turkeys, Eimeria species, Ornithobacterium rhinotracheale, Pasteurella multocida, Mycoplasma synoviae, Salmonella species and E. coli.
- 52) Method for the preparation of a vaccine according to claims 47-51, said method comprising the admixing of a nucleic acid according to claims 1-16, a DNA fragment according to claim 17, a recombinant DNA molecule according to claim 18, a live recombinant carrier according to claim 19, a host cell according to claim 20, a protein according to claims 21-44 or an immunogenic fragment thereof, or antibodies against a protein according to claims 21-44 and a pharmaceutically acceptable carrier.

Abstract

The present invention relates to nucleic acids encoding *Ornithobacterium rhinotracheale* proteins, to DNA fragments, recombinant DNA molecules, live recombinant carriers and to host cells comprising such nucleic acids. The present invention also relates to *Ornithobacterium rhinotracheale* proteins and to antibodies against such proteins. Another embodiment of the invention relates to such proteins for use in vaccines and to the use of such proteins in the manufacturing of such vaccines. Also an embodiment of the invention relates to vaccines comprising such nucleic acids, DNA fragments, recombinant DNA molecules, live recombinant carriers, host cells, proteins or antibodies against such proteins. Finally, again another embodiment of the invention relates to methods for the preparation of such vaccines.

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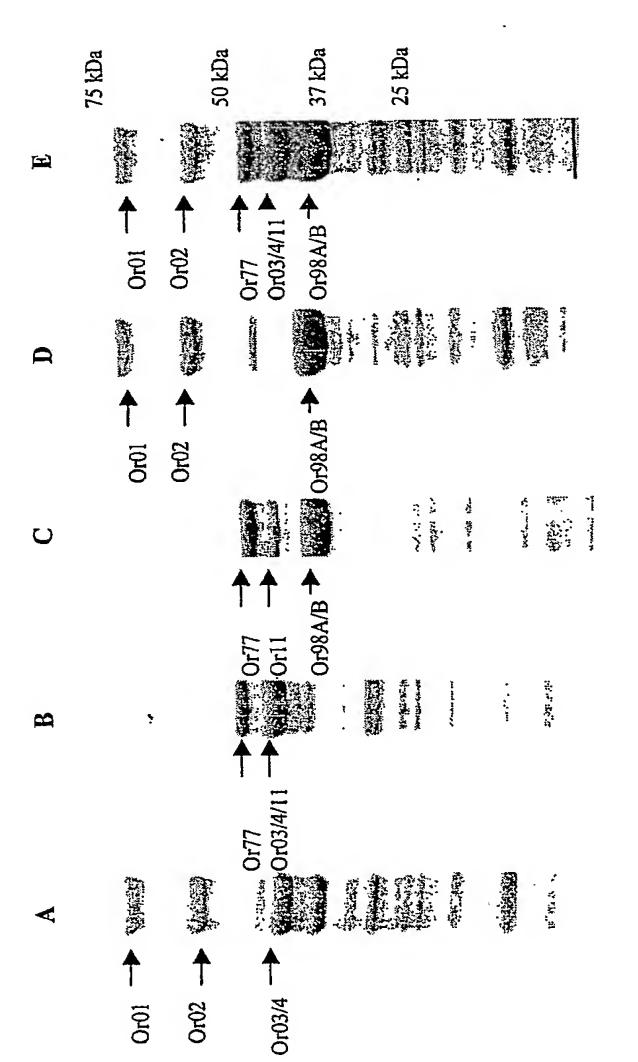


Figure 1.

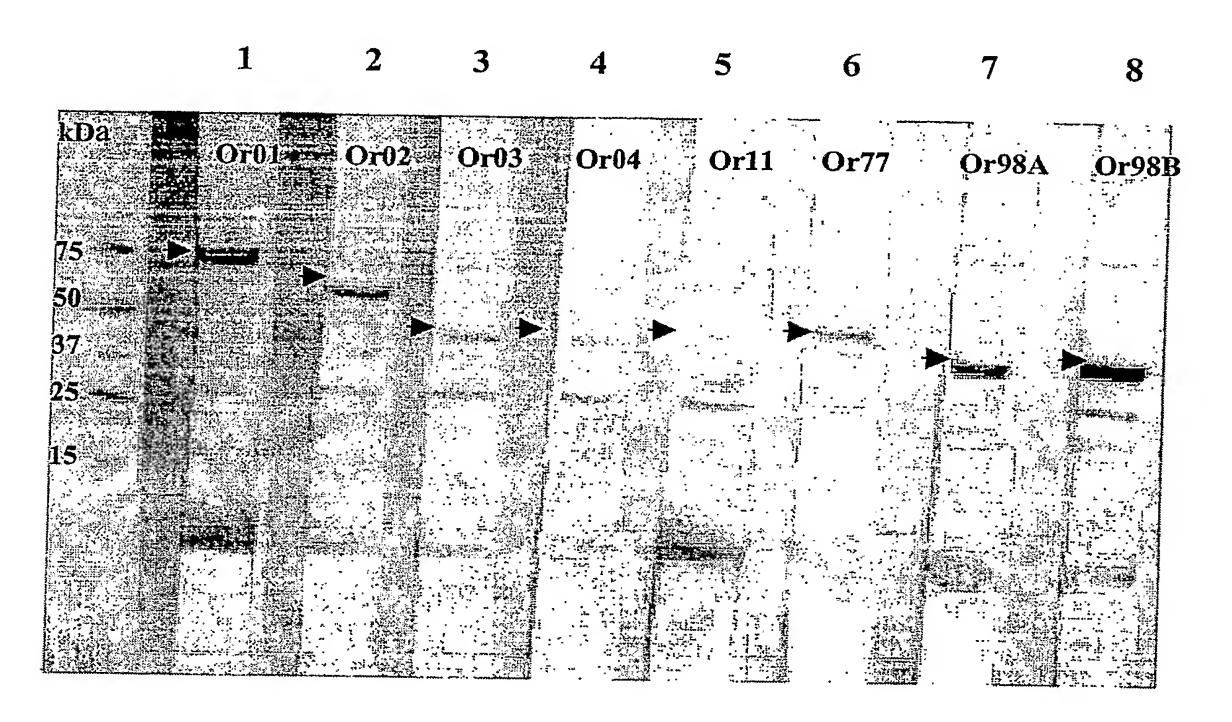
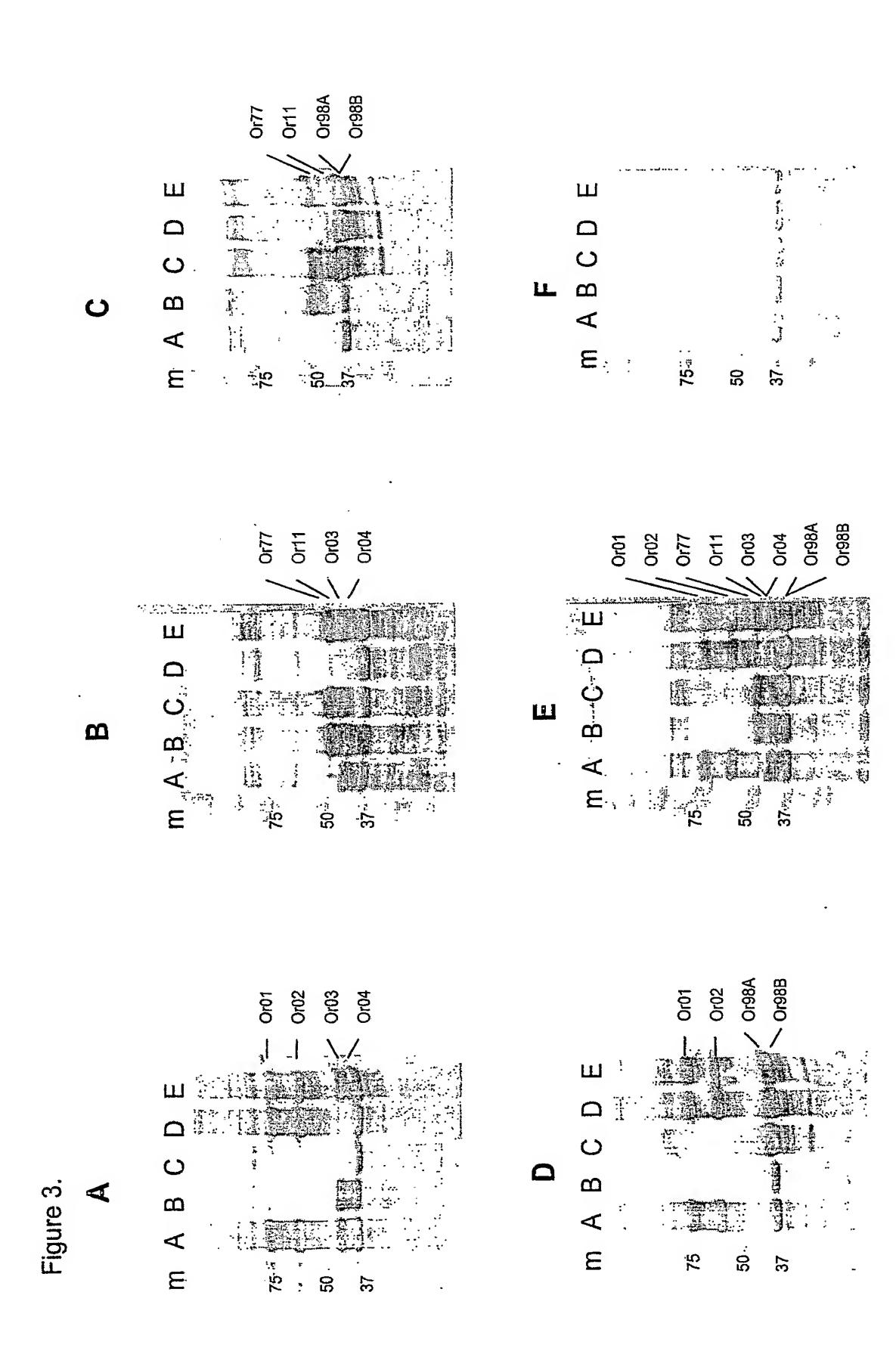


Figure 2.

Strip	Protein on blot:	Sera from birds vaccinated with:
1	Or01	Or01
2	Or02	Or02
3	Or03	Or03
4	Or04	Or04
5	Or11	Or11
6	Or77	Or77
7	Or98A	Or98A
8	Or98B	Or98B



Cross-protection subunit vaccination

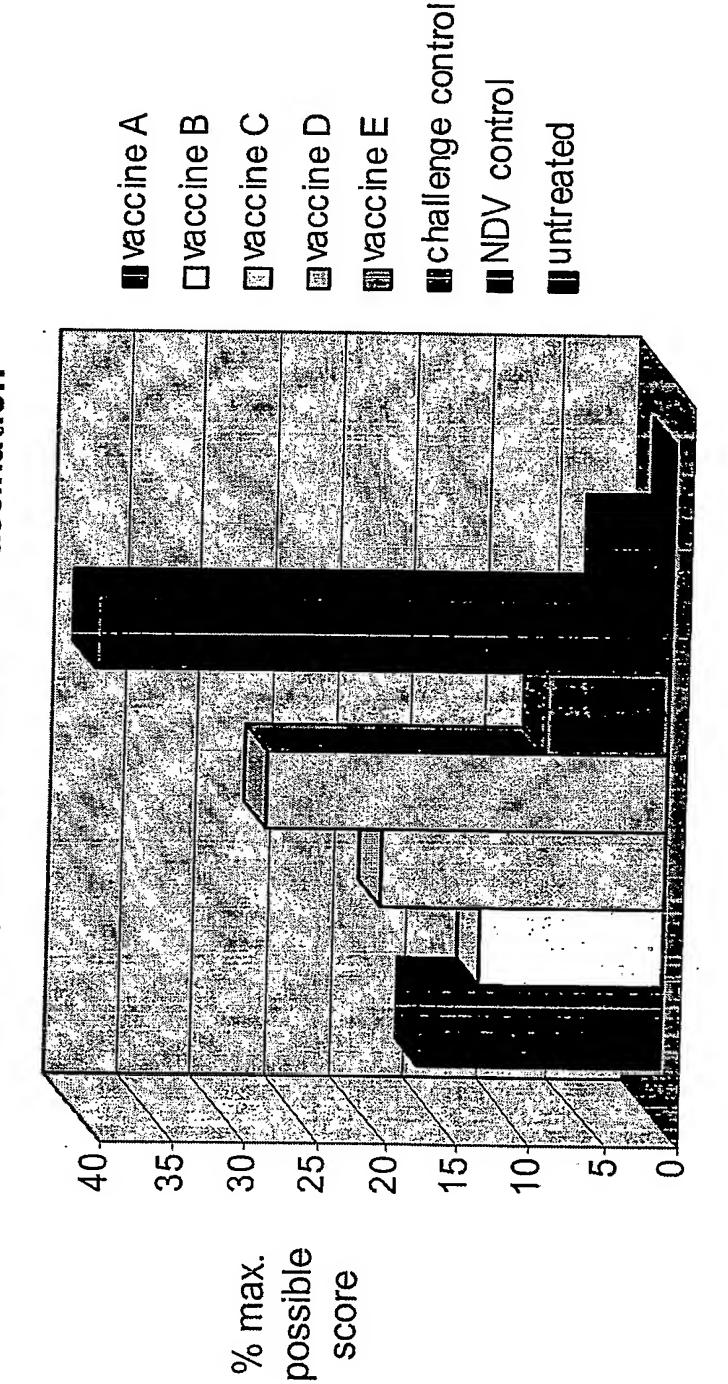


Figure 4.

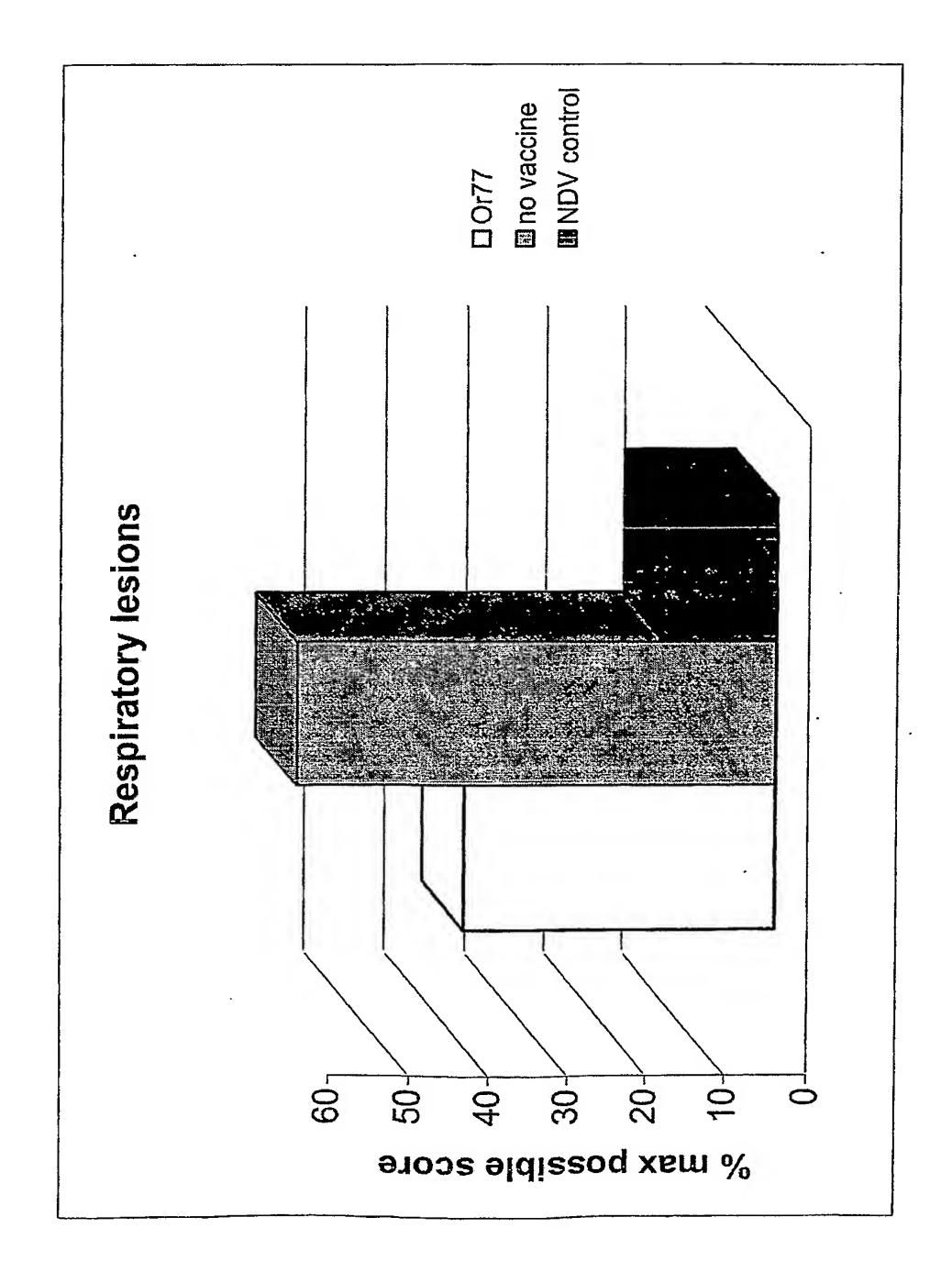
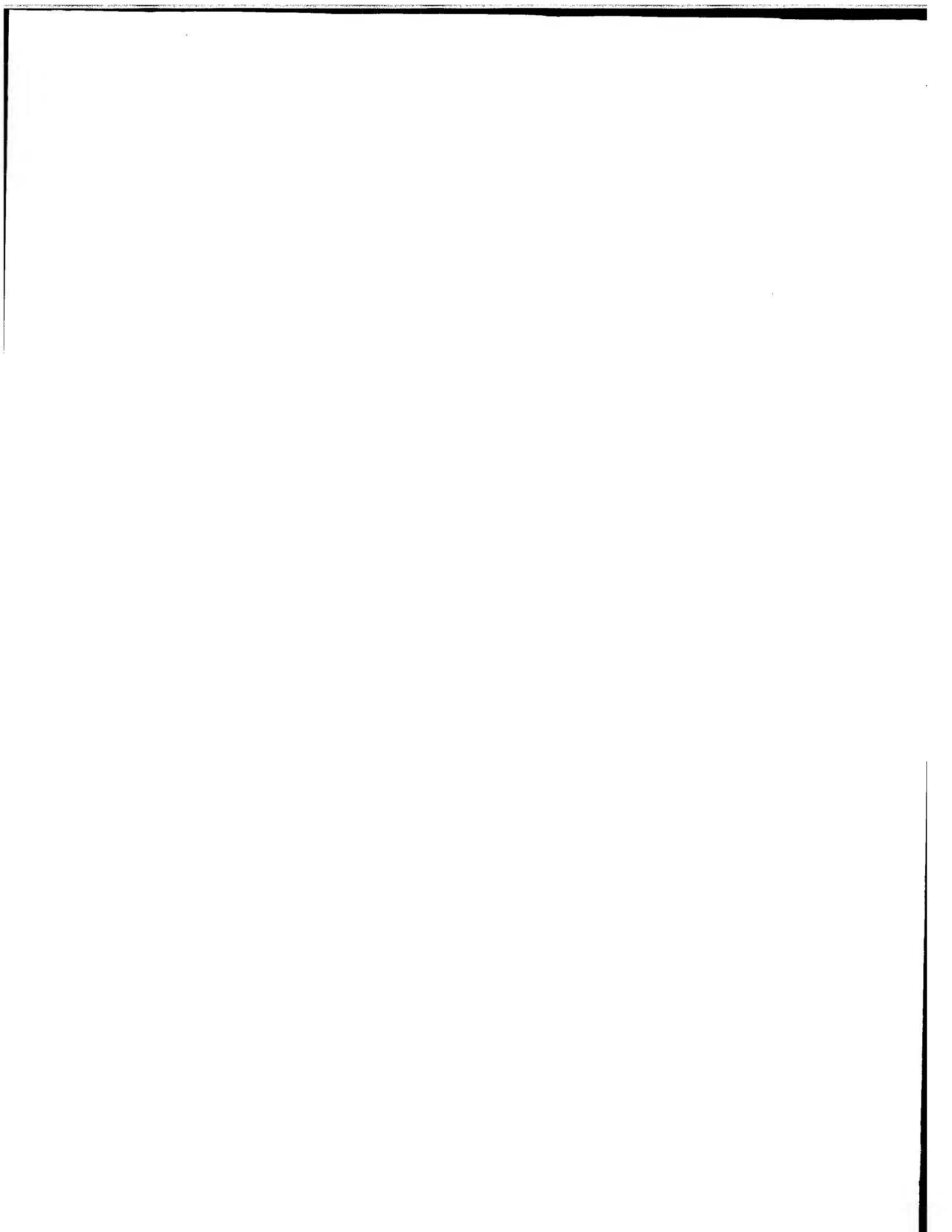


Figure 5.



SEQUENCE LISTING

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Val			Ala	Thr	Gly	Ser		Phe	Leu	Gln	Thr	Leu	Lys	Gln	Tyr	
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Gly Lys Val Glu Ser Trp Asn Lys Lys Val Gly Asp Lys Val Ser Tyr

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Gln Ala Ala Pro Val Asp Ser Ile Leu Ala Ile Ile Gly Ala Glu Gly 65 70 75 80

Glu Asp Ile Ser Gly Leu Val Ser Gly Gly Gly Ala Ser Gln Ser Ala 85 90 95

Pro Ala Glu Glu Ala Ala Ala Pro Ala Glu Glu Pro Glu Ala Glu Ala 100 105 110

Ala Pro Ala Ala Glu Val Pro Glu Asn Val Thr Ile Val Ser Met Pro 115 · 120 125

Arg Leu Ser Asp Thr Met Glu Glu Gly Lys Val Glu Ser Trp Asn Lys
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Lys Val Gly Asp Lys Val Ser Tyr Gly Asp Ile Leu Ala Glu Ile Glu
145 150 155 160

Thr Asp Lys Ala Val Gln Glu Phe Glu Thr Asp Val Glu Gly Thr Leu 165 170 175

Leu Tyr Ile Gly Val Glu Ala Gly Gln Ser Ala Pro Val Asp Ser Ile 180 185 190

Leu Ala Ile Ile Gly Pro Glu Gly Thr Asp Val Ser Ala Ile Val Ala 195 200 205

Gly Gly Ala Lys Pro Ala Ala Lys Ala Glu Ala Pro Lys Ala Glu 210 215 220

Ala Pro Lys Gln Ala Ala Pro Ala Gln Glu Lys Lys Glu Thr Pro Ala

225

230

235

240

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Ile Ser Pro Leu Ala Lys Lys Leu Ala Asp Glu Lys Gly Tyr Asp Ile 260 265 270

Asn Gln Ile Gln Gly Thr Gly Asp Asn Gly Arg Ile Ile Lys Lys Asp 275 280 285

Val Glu Asn Phe Thr Pro Gln Ala Ala Ala Ala Lys Pro Ala Val Ala 290 295 300

Gly Pro Val Ala Leu Glu Val Gly Glu Asp Thr Val Ile Pro Asn Ser 305 310 315 320

Gln Met Arg Lys Val Ile Ala Lys Arg Leu Ser Glu Ser Lys Phe Thr
325 330 335

Ala Pro His Tyr Tyr Leu Thr Ile Glu Val Asp Met Asp Asn Val Met 340 345 350

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Val Val Asn Ser Thr Trp Lys Asp Asn Glu Ile Val Gln Tyr Ala Ala 385 390 395 400

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Val Val Lys Asn Thr Asp Leu Lys Ser Leu Ser Gln Ile Ser Ala Glu
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Val Lys Asp Leu Ala Thr Arg Ser Arg Asp Arg Lys Ile Lys Ala Asp 435 440 445

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Val Glu Ser Phe Thr Ser Ile Ile Asn Gln Pro Asn Ser Cys Ile Leu 465 470 475 480

Ser Val Gly Ala Ile Val Glu Lys Pro Val Val Lys Asn Gly Gln Ile 485 490 495

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7.11	T Try			GTU	r WT	A AS	b bř			Lys	Ty:	r Va	L P	Ala		ı Gl	у А	sn	Ser	
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Le	u Th	r S	Ser	Gly	Туг	: Sei	r As	p G	Ly F	lla	Let	ı Ty	r A	rg	Ser	Al	a G	ln	Glu	
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Val	PLU	GT.	y A	ידים י			ryr	HLS	re	u i			GL:	n G	31y	Tyr	Gly	y A	.sn	
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355 360 365

tta ttg cca ctt aca gcg agt aga aca ctt ggg aaa tta aat agt gaa Leu Leu Pro Leu Thr Ala Ser Arg Thr Leu Gly Lys Leu Asn Ser Glu aga ctt gct act ttg aca aaa tta gga tta cca aag gaa aac gcc gct Arg Leu Ala Thr Leu Thr Lys Leu Gly Leu Pro Lys Glu Asn Ala Ala caa ctt tct atg aac gga ctt act tat cca ttg caa gat gcc gat gtt Gln Leu Ser Met Asn Gly Leu Thr Tyr Pro Leu Gln Asp Ala Asp Val tta acc aaa aat gaa gtt tca aca att cac gaa aga gta aac gaa atc Leu Thr Lys Asn Glu Val Ser Thr Ile His Glu Arg Val Asn Glu Ile aat caa ggc ata caa gca gtg gca aaa caa ttc aac att gca tat gtg Asn Gln Gly Ile Gln Ala Val Ala Lys Gln Phe Asn Ile Ala Tyr Val gac atg aat gcc gaa atg caa aaa ctc act aaa ggc ttt aaa ttc aac Asp Met Asn Ala Glu Met Gln Lys Leu Thr Lys Gly Phe Lys Phe Asn Gly Val Asp Tyr Asn Ala Ser Phe Val Thr Gly Gly Ala Phe Ser Leu gat gga gtg cat tta aac agc cga gga tat gcc cat aca gct aat aca Asp Gly Val His Leu Asn Ser Arg Gly Tyr Ala His Thr Ala Asn Thr ttt att cgt gcc atc aat cag caa tat aag gca agc att ccg ttg gta Phe Ile Arg Ala Ile Asn Gln Gln Tyr Lys Ala Ser Ile Pro Leu Val gat atc aac gct ttc cca ggc aca caa tta cct taa Asp Ile Asn Ala Phe Pro Gly Thr Gln Leu Pro 520 ·

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Leu Thr Ser Gly Tyr Ser Asp Gly Ala Leu Tyr Arg Ser Ala Gln Glu 50 55 60

Asn Ser Tyr Pro Ala Ile Ile Ala Lys Gln Met Lys Tyr Val Gly Gly 65 70 75 80

Gly Glu Phe Ser Gln Pro Leu Met Lys Asp Asn Ile Gly Gly Phe Ser 85 90 95

Asp Leu Phe Glu Ala Ser Lys His Thr Ala Phe Tyr Gly Lys Leu Glu 100 105 110

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Phe Ser Leu Ala Gln Thr Phe Val Lys Gly Asn Phe Asn Asn Leu Gly 130

Val Pro Gly Ala Lys Ser Tyr His Leu Leu Ala Gln Gly Tyr Gly Asn 145 150 155 160 Ile Ala Asn Leu Lys Glu Ser Lys Ala Asn Pro Tyr Phe Val Arg Phe
165 170 175

Ala Ser Gln Pro Asn Ala Ser Val Leu Ser Asp Ala Leu Ala Gln Lys
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Pro Thr Phe Phe Thr Leu Trp Ile Gly Asn Asn Asp Val Leu Gly Tyr
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Ser Ile Gln Lys Leu Val Lys Ala Leu Thr Asp Ser Gly Ala Lys Gly
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Val Pro Ala Glu Pro Leu Ser Pro Leu Asn Lys Ser Tyr Ala Thr Gln 275 280 285

Ile Glu Asn Leu Asn Lys Phe Tyr Ala Ser Leu Asn Lys Val Phe Asp 290 295 300

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Ser Gly Ala Val Ile Val Asp Lys Ser Leu Pro Asp Leu Ser Gln Lys
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330
335

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340 345 350

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Arg Leu Ala Thr Leu Thr Lys Leu Gly Leu Pro Lys Glu Asn Ala Ala 385

Gln Leu Ser Met Asn Gly Leu Thr Tyr Pro Leu Gln Asp Ala Asp Val 405 410 415

Leu Thr Lys Asn Glu Val Ser Thr Ile His Glu Arg Val Asn Glu Ile 420 . 430

Asn Gln Gly Ile Gln Ala Val Ala Lys Gln Phe Asn Ile Ala Tyr Val 435 440 445

Asp Met Asn Ala Glu Met Gln Lys Leu Thr Lys Gly Phe Lys Phe Asn 450 455 460

Gly Val Asp Tyr Asn Ala Ser Phe Val Thr Gly Gly Ala Phe Ser Leu 475 480

Asp Gly Val His Leu Asn Ser Arg Gly Tyr Ala His Thr Ala Asn Thr 485 490 495

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Tyr	Gly	Gly	Leu	Ile	Phe	Asp	Glu	Ser		Gln	Thr	Leu	Lys		Glu	
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				-										s. h= 3		03.6
		_				aat										816
Asp	Gly	Val		Val	Ile	Asn	Leu		Phe	Val	Pro	TTG		Pne	гÀг	
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Lys Arg Gly Asp Phe Glu Phe Leu Ile Leu Lys Lys Ser Asn Asp Thr 130 135 140

Leu Tyr Leu Lys Gly Lys Lys Thr Gly Asn Tyr Met Lys Leu Tyr Lys 145 150 155 160

Ala Gly Asn Ile Gln Glu Ile Lys Ser Asn Ile Arg Lys Val Ala Thr 165 170 175

Thr Ile Asp Arg Val Asp Leu Pro Ala Gln Gly Thr Ile Gly Thr Glu 180 185 190

Pro Leu Val Leu Ser Thr Gly Gly Thr Arg Asn Ile Ile Phe Ser Thr 195 200 205

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Asp Gly Val Ile Val Ile Asn Leu Lys Phe Val Pro Ile Asn Phe Lys 260 265 270

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Gly Tyr Lys Lys Ala Arg Ala Gly Asp Ser Leu Leu His Gly Met Ile 290 295 300

Leu Ser Lys Phe Lys Leu Gln Asp Phe Tyr Val Leu Gly Asn Phe Arg 305

Asp Asn Val Gly Phe Asn Thr Phe Val Glu Gly Tyr Asn Gly Ala Phe 325 330 335

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Asp	Ser	Phe	Thr	Pro	Ala	Pro	Pro	Thr	Glu	Lys	Lys		Asp	Thr	Pro		
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Leu	Ile	Asn	Leu	Leu	Asp	Asp	Phe	Val	Phe	Phe		Lys	Asp	Val	Val		
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Thr	Ile	Pro	Val	Asp	Lys	Asp	Asn	Leu	Ala		Asn	Asn	vaı	TTE			
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Gly	Glu	Val	Phe		Asn	Arg	Lys	Met		Glu	Asn	Pne	GLU	1yr 95	GIII		
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					gat											•	.
Leu	Glu	Leu		GIn	Asp	.i.rb	тте		ser	ASN	5 T.O	wsb		TILL	ಬ್ಗಡ		
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					ttt											•	J U 4
Ile	Pro	Asn	GTA	ALa	Phe	Thr	тте	ser	GTÀ	GIN	IUL	ьеu	wan	ыXы	120F)		

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Lys Lys Giy Tyr			sn Ile Pro Arg Thr	
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210	215		20	
			ca gac act gat gat	720
		a Asn Trp Trp Al	la Asp Thr Asp Asp	
225	230	235	240	
aac aca aca tat	Ctt dat dta aaa	a tto oot att aa	at aca ata aaa gct	
			on Thr Ile Lys Ala	768
	245	250	255	
•			202	
ata aaa tta tac	act aaa agc tat	tgg caa aat gc	t gta ggc agt gta	816
			a Val Gly Ser Val	
260		265	270	
		ggc aat act tgg	g aaa gaa cag gga	864
The Transfer Agr (O N N	63	p Lys Glu Gln Gly	

att gct aac ttt ggg caa tat tca aca gtg tct act att gta ttc act

Ile Ala Asn Phe Gly Gln Tyr Ser Thr Val Ser Thr Ile Val Phe Thr

caa cca att gac att aat gct gtc aga ata tct aac ttc act aga ggg 960 Gln Pro Ile Asp Ile Asn Ala Val Arg Ile Ser Asn Phe Thr Arg Gly 320 315 310 305 gga agt agt aat ttc att aac att aac gag gtg gaa gta ttc aaa ata 1008 Gly Ser Ser Asn Phe Ile Asn Ile Asn Glu Val Glu Val Phe Lys Ile 335 330 325 1023 cca agt gaa gaa taa Pro Ser Glu Glu 340 <210> 8 <211> 340 <212> PRT <213> Ornithobacterium rhinotracheale : <400> 8 Met Lys Asp Ile Phe Glu Tyr Thr Leu Leu Ala Leu Gly Gly Leu Leu 15 10 5 1 Leu Thr Asn Cys Tyr Asp Ser Asp Glu Ile Glu Val Ile Lys Phe Asp 30 25 20 Asp Ser Phe Thr Pro Ala Pro Pro Thr Glu Lys Lys Arg Asp Thr Pro 45 40 35 Leu Ile Asn Leu Leu Asp Asp Phe Val Phe Phe Lys Lys Asp Val Val

Thr Ile Pro Val Asp Lys Asp Asn Leu Ala Thr Asn Asn Val Ile Ser 65 70 75 80

55

50

60

Gly Glu Val Phe Thr Asn Arg Lys Met Ser Glu Asn Phe Glu Tyr Gln 85 90 95

Leu Glu Leu Asp Gln Asp Trp Ile Ser Ser Asn Pro Asp Leu Gln Ala 100 105 110

Ile Pro Asn Gly Ala Phe Thr Ile Ser Gly Gln Thr Leu Asn Lys Asp 115 120 125

Glu Arg Asn Gly Thr Phe Lys Ile Gln Leu Asn Ala Glu Val Ala Lys 130 135 140

Glu Leu Gly Gly Thr Tyr Tyr Leu Pro Leu Lys Leu Val Ser Lys Asn 145 150 155 160

Asp Asn Leu Asn Ile Leu Lys Gly Tyr Glu Ser Gly Val Phe Lys Leu 165 · 170 175

Val Phe Lys Lys Ser Tyr Pro Ile Pro Glu Gly Asn Asn Val Glu Gly
180 185 190

Lys Lys Gly Tyr Tyr Phe Asp Gly Leu Gly Asn Asn Ile Pro Arg Thr
195 200 205

Asp Leu Ser Phe Asn Ser Asn Tyr Ala Pro Asp His Leu Phe Lys Leu 210 225 220

Asn Asp Gly Asn Gln Gln Gly Ala Asn Trp Trp Ala Asp Thr Asp Asp 225 230 235 240

Asn Thr Tyr Leu Asp Val Lys Phe Pro Ile Asn Thr Ile Lys Ala
245 250 255

Ile Lys Leu Tyr Thr Lys Ser Tyr Trp Gln Asn Ala Val Gly Ser Val
260 265 270

Lys Ile Glu Val Ser Asn Asp Asn Gly Asn Thr Trp Lys Glu Gln Gly

275 280 285

Ile Ala Asn Phe Gly Gln Tyr Ser Thr Val Ser Thr Ile Val Phe Thr 290 295 300

Gln Pro Ile Asp Ile Asn Ala Val Arg Ile Ser Asn Phe Thr Arg Gly 305 310 315 320

Gly Ser Ser Asn Phe Ile Asn Ile Asn Glu Val Glu Val Phe Lys Ile 325 330 335

Pro Ser Glu Glu 340

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<220>

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48

att tta tgg gca ggc gga tac cga gtt tcg ctg caa ggt gta aga caa 96

Ile Leu Trp Ala Gly Gly Tyr Arg Val Ser Leu Gln Gly Val Arg Gln
20 25 30

gcc gcc atg ggg gca caa ggt gta gca ctt tct cac gat gcg agt gtg
Ala Ala Met Gly Ala Gln Gly Val Ala Leu Ser His Asp Ala Ser Val

40
45

gca ttt ttc aac ccc gca gca ttg gct ttt gta gat gat aaa tta agt

Ala Phe Phe Asn Pro Ala Ala Leu Ala Phe Val Asp Asp Lys Leu Ser

50 55 60

																		aac	
65	: AL	a v	ar G	тА	3T.À	Phe	G17	7 Il	e Gl	Ly I	le	Thr 75	Ala	а Ьу	rs T	'yr	Glr	Asn	L
						. •						, ,						80	
																		cca	
Arg	G1	u Th	r L			Lys	Ala	Gl	u Th			Asn	Pro	Le	u G	ly	Thr	Pro	
				8	5					9	0						95		
ctt	tai	c ct	t go	ct a	ca a	agc	tat	aag	g cc	t a	cg	gaa	aaa	ct	a go	2C	tta	ggc	336
																		Gly	
			10	0					10	5					13	LO			
gtg	ago	: at	a ac	c a	at d	eca	tet	aac	i aci	r ac	יר ו	c ta	asa	t-00	· ~~	•	~~ h		224
Val																			384
		11						120						125		_	-	-	
too	aat	~~				. 4													
tgg Trp																			432
	130						135			,			140	my s	, ne	ا. خاد،	FIIE	FILE	
att d																		-	480
Ile (145	GT11	ET () <u>1</u> 11.	C Al		ла 50	ryr	гла	vaı	. Tn		.55	Trp	Leu	Se	r (Gly 160	
											_							700	
gct s																			528
Ala (Зlу	Ala	ı Ile	: Il 16		la A	Arg	Gly	Asn			sn	Ile	Lys	Arg			Ile	
				Τ.0	J					170	U					1	75		
tct c	ta	ggc	aac	ca	a ga	at ç	gcg (999	cta	gaa	a a	tc s	gac	aaa	aaa	ı g	ga g	gct	576
Ser I	eu	Gly	Ası	Gl:	n As	sp F	Mla	Gly	Leu	Glı	ıI	le A	Asp	Lys	Lys	G	ly A	Ala	
			180	•					185						190)			
cac g	ga	aca	999	tt	: aa	at g	ıta ç	399	gtt	tat	: q	cc a	aaa (cca	aat	a:	at a	naa	624
His G																			024
		195					3	200					:	205					
tta a	at	ata	aaa	att	- ດຕ	1 4	ac (7072	tas	~~~	~ •			- b					
tta a Leu A																			672
	10						- 15	_					20	-	— <u>,</u>	~ ~ ~	- ~ 11	υp	
aaa g																			720
Lys Gl 225	- A 1	≁≈ħ	нта	val	23		ys A	sn]	Leu	Pro	\$e 23		le V	al :	Lys	Gl		_	
						=					دے	-					2	40	

atg	cct	ttt	tcg	gct	aaa	tat	ttt	gat	gct	caa	tta	cct	cta	cca	gca	768
Met	Pro	Phe	Ser	Ala	Lys	Tyr	Phe	Asp	Ala	Gln	Leu	Pro	Leu	Pro	Ala	
				245					250					255		
gaa	ctt	tta	att	999	gcg	aac	tat	aaa	gta	aca	cca	aaa	ttg	ctc	gta	816
	Leu															
			260					265					270			
999	gca	qaa	att	999	gct	gta	aaa	tgg	aac	gcc	tac	gaa	aca	tta	aat	864
	Ala															
-		275					280					285				
att	aaa	ctt	tat	aac	aac	gaa	gag	gaa	tac	aac	aat	act	tct	aac	aaa	912
	Lys															
	290					295					300					
aat	tac	aaa	aac	aca	tta	aat	tat	agt	atc	999	gct	gaa	tat	tta	atc	960
	Tyr															
305					310		_			315					320	
aat	cca	aaa	act	qcc	tta	cqc	tta	999	tat	aaa	ttc	gac	aaa	tcg	cct	1008
	Pro															
				325		J		_	330	_				335		
tca	cca	act	gat	tcq	ttt	aac	cca	gag	acc	cca	acc	att	aat	tat	cac	1056
_	Pro															
			340					345					350			
qca	ttt	aca	act	gga	ttt	gga	tat	gaa	ttc	gag	aga	ttt	cgt	gta	gat	1104
_	Phe															
		355		_		_	360					365				
			•											•		
qcc	atg	qcq	gaa	tat	tta	cta	gga	aac	gaa	aga	agc	ttc	cac	aat	aca	1152
	Met	·														
	370			-		375					380					
caa	tat	aac	ttt	999	ggc	gac	atc	aac	act	ggt	ggc	tat	gtg	ttt	ggt	1200
	Tyr															
385	_				390	_				395					400	
								•								
cta	ggt	tta	tca	tat	aga	ctt	gac	aaa	taa							1230
	Gly															
~~	<u> </u>			405			<u>.</u>	•								

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<211> 409

<212> PRT

<213> Ornithobacterium rhinotracheale

<400> 10

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1 10 15

Ile Leu Trp Ala Gly Gly Tyr Arg Val Ser Leu Gln Gly Val Arg Gln
20 25 30

Ala Ala Met Gly Ala Gln Gly Val Ala Leu Ser His Asp Ala Ser Val $4\dot{0}$ 45

Ala Phe Phe Asn Pro Ala Ala Leu Ala Phe Val Asp Asp Lys Leu Ser 50 55 60

Ile Ala Val Gly Gly Phe Gly Ile Gly Ile Thr Ala Lys Tyr Gln Asn 70 75 80

Arg Glu Thr Leu Tyr Lys Ala Glu Thr Asp Asn Pro Leu Gly Thr Pro
85 90 95

Leu Tyr Leu Ala Thr Ser Tyr Lys Pro Thr Glu Lys Leu Ala Leu Gly
100 105 110

Val Ser Val Thr Thr Pro Phe Gly Ser Thr Val Asp Trp Gly Asp Lys
115 120 125

Trp Ala Gly Arg Tyr Ile Ile Asp Arg Ile Ala Leu Lys Ser Phe Phe 130 135 140

Ile Gln Pro Thr Ala Ala Tyr Lys Val Thr Asp Trp Leu Ser Val Gly
145 150 155 160

Ala Gly Ala Ile Ile Ala Arg Gly Asn Val Asn Ile Lys Arg Ala Ile 165 170 175

Ser Leu Gly Asn Gln Asp Ala Gly Leu Glu Ile Asp Lys Lys Gly Ala 180 185. 190

His Gly Thr Gly Phe Asn Val Gly Val Tyr Ala Lys Pro Asn Asp Lys
195 200 205

Leu Asn Ile Gly Ile Ala Tyr Arg Ser Glu Val Lys Met Lys Ala Asp 210 215 220

Lys Gly Asp Ala Val Phe Lys Asn Leu Pro Ser Ile Val Lys Gly Lys 225 230 230 235 235

Met Pro Phe Ser Ala Lys Tyr Phe Asp Ala Gln Leu Pro Leu Pro Ala 255

Glu Leu Leu Ile Gly Ala Asn Tyr Lys Val Thr Pro Lys Leu Leu Val 260 265 270

Gly Ala Glu Ile Gly Ala Val Lys Trp Asn Ala Tyr Glu Thr Leu Asn 275 280 285

Ile Lys Leu Tyr Asn Asn Glu Glu Glu Tyr Asn Asn Thr Ser Asn Lys 290 295 300

Asn Tyr Lys Asn Thr Leu Asn Tyr Ser Ile Gly Ala Glu Tyr Leu Ile 305 310 310 320

Asn Pro Lys Ala Ala Leu Arg Leu Gly Tyr Lys Phe Asp Lys Ser Pro 325 330 335

Ser Pro	Ala Asp Ser Phe 340	Asn Pro Glu 345	Thr Pro Thr Ile	e Asn Tyr His 350
	Thr Thr Gly Phe	Gly Tyr Glu 360	Phe Glu Arg Phe	
Ala Met A	da Glu Tyr Leu	Leu Gly Asn 375	Glu Arg Ser Phe	His Asn Thr
Gln Tyr A 385	sn Phe Gly Gly 390	Asp Ile Asn	Thr Gly Gly Tyr 395	Val Phe Gly 400
Leu Gly L	eu Ser Tyr Arg 405	Leu Asp Lys	•	
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		Ala Ile Ser F	tt tcg tct ttt he Ser Ser Phe	•
			ca cct aaa gaa a hr Pro Lys Glu 1	_
			ag cca gat gaa d ys Pro Asp Glu E . 45	

gat gat gga aac gaa aat cca gaa aac act gga gat gaa gag aat gga

Asp	Asp 50	Gly	Asn	Glu	Pro 55	Glu	Asn	Thr	Gly	Asp 60	Glu	Glu	Asn	Gly		
	aat Asn														240	
	cgc Arg														288	
	gat Asp														336	
	gct Ala														384	
	aac Asn 130														432	
	aaa Lys														480	
	agc Ser														528	\$ *.*
	ttg Leu		Asn	ttt				atc							576	
	gca Ala	Ser				Gly	att					gga			624	
	Glå aaa									Tyr	aat				672	
	GJÅ 333 510														720	

225					230	0				23!	ō				240	
) Lys					: Ile					aaa Lys	
				Phe					: Ser					Leu	tcc Ser	816
att ç Ile A			Thr										Lys			864
aga a Arg A 2																912
aac g Asn G 305																960
gaa t																1008
ata go							Leu					Lys				1.056
tgg tt	eu 1					Lys					Asn (1104
gat tt Asp Le 37	u V				Leu A						caa					1140
<210><211><211><212><213>	12 37 PR	9 T	ıobac	cteri	lum r	chinc	otrac	hea]	le							

<400> 12

Met Lys Lys Tle Leu Leu Ala Ile Ser Phe Ser Ser Phe Val Leu Ser 1 5 10 15

Cys Ser Ser Asp Asp Tyr Thr Pro Ala Thr Pro Lys Glu Thr Glu Lys
20 25 30

Pro Lys Glu Glu Ala Val Val Pro Asn Lys Pro Asp Glu Pro Lys Ala 35 40 45

Asp Asp Gly Asn Glu Asn Pro Glu Asn Thr Gly Asp Glu Glu Asn Gly 50 55 60

Asp Asn Thr Asn Ser Val Val Gly Lys Pro Asp Asp Phe His Met Gly 65 70 . 75 80

Asn Arg Ser Tyr Ala Ser Trp Lys Glu Asp Val Asp Tyr Ile Gly Gly 85 90 95

Phe Asp Ile Glu Thr Leu Leu Ser Gly Ala Asp Asn Gln Lys Tyr Asp 100 105 110

Ala Ala Tyr Phe Ser Gln Phe Ile Lys Ile Phe Ser Ser Pro Asn 115 120 125

Gly Asn Asn Phe Tyr Thr Phe Gln Ala Glu Asp Phe Lys Asp Val Glu 130 135 140

Thr Ser Tyr Lys Gly Val Lys Ser Glu Ile Thr Ser Ser Leu Lys Phe 165 170 175 Asp Leu Ala Asn Phe Tyr Asp Arg Lys Ile Lys Ile Asn Glu Asp Phe 180 185 190

Val Ala Ser His Tyr Met Arg Gly Ile Tyr Glu Glu Leu Gly Gly Phe 195 200 205

Ile Gly Asn Leu Leu Asn Tyr Asp Asp Glu Lys Tyr Asn Leu Glu Leu 210 215 220

Ala Gly Ser Lys Asn Lys Asp Glu Ser Asn Asn Ser Leu Gly Phe Ser 225 230 235 240

Ile Arg Val Thr Asp Lys Lys Asp Lys Tyr Ile Thr Thr Val Tyr Lys
245 250 255

Asn Ile Ser Gly Phe Arg Pro Leu Ser Ser Leu Gln Glu Glu Leu Ser 260 265 270

Ile Ala Pro Thr Tyr Glu Leu Arg Glu Lys Ile Lys Glu Lys Ile Asp ·275 280 285

Arg Asn Lys Arg Asn Ile Ser Leu Leu Glu Leu Leu Lys Pro Ser Val 290 295 300

Asn Glu Trp Met Lys Ser Ala Asp Phe Tyr Phe Asn Asn Thr Asp Leu 305 310 315 320

Glu Trp Arg Gly Asp His Tyr Ser Ala Arg Gly Phe Leu Asp Leu Tyr
325 330 335

Ile Gly Ser Pro Arg Phe Glu Leu Ile Leu Ala Thr Lys Glu Asp Asn 340 345 350

Trp Leu Ile Leu Lys Val Lys Val Val Gln Ile Asn Glu Val Pro Thr

355 360 365

Asp Leu Val Tyr Ser Leu Arg Val Ser Ile Asn 370 375

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ggt aag tta atg gtt ggt gtc aag cca cca tta acc cct aat caa gag

Gly Lys Leu Met Val Gly Val Lys Pro Pro Leu Thr Pro Asn Gln Glu

20 25 30

48

aag ttg ctc aca gac tta gag ggc aaa atg gaa gct ggg acc att acc
Lys Leu Leu Thr Asp Leu Glu Gly Lys Met Glu Ala Gly Thr Ile Thr

40
45

aaa aag caa atc atc act tat ggt gaa ttg ctt tcc aag aaa aac caa 192 Lys Lys Gln Ile Ile Thr Tyr Gly Glu Leu Leu Ser Lys Lys Asn Gln 50

aag ctt gaa tta tct gca agt gta aag tct tac tta gcc gac att cat
Lys Leu Glu Leu Ser Ala Ser Val Lys Ser Tyr Leu Ala Asp Ile His

70 75 80

aaa gaa gtc ttt ttt ggt cgt gat aag gaa ttg acc aat aaa tat cta
Lys Glu Val Phe Phe Gly Arg Asp Lys Glu Leu Thr Asn Lys Tyr Leu
85 90 95

tca aaa ggc att caa gta gaa gaa aag agc ata acg ctc tat tcc gat

Ser Lys Gly Ile Gln Val Glu Glu Lys Ser Ile Thr Leu Tyr Ser Asp

100 105 110

gtc tgt aac aag tta ttc cta aag aat aaa aag ttt tac aaa aac gat Val Cys Asn Lys Leu Phe Leu Lys Asn Lys Lys Phe Tyr Lys Asn Asp 115 120 125	384
ttt att caa ggt acg cca gat aac acg caa gac aaa atc aga gat atc Phe Ile Gln Gly Thr Pro Asp Asn Thr Gln Asp Lys Ile Arg Asp Ile 130 135 140	432
aaa agt agt tgg gac ttc tca acc ttt cct cta cac gcc gat gaa acg Lys Ser Ser Trp Asp Phe Ser Thr Phe Pro Leu His Ala Asp Glu Thr 145 150 155 160	480
cca acc aaa gac tat gaa tgg cag ttg caa ggt tat atg gaa tta aca Pro Thr Lys Asp Tyr Glu Trp Gln Leu Gln Gly Tyr Met Glu Leu Thr 165 170 175	528
ggc tta aaa gaa gct gag ttg att tat tgc ttg gtt gat acg cct·cat Gly Leu Lys Glu Ala Glu Leu Ile Tyr Cys Leu Val Asp Thr Pro His 180 185 190	576
aaa att gta gaa gat gaa atc cga aga atg gac tgg aag cat aat tta Lys Ile Val Glu Asp Glu Ile Arg Arg Met Asp Trp Lys His Asn Leu 195 200 205	624
ctt gac att aac ggc gaa gtg aga gcc gag aca aga gat tta gta gtt Leu Asp Ile Asn Gly Glu Val Arg Ala Glu Thr Arg Asp Leu Val Val 210 215 220	672
gag att gtg tct aac tta att tat acc aag caa ggc ttg gaa gac ttt Glu Ile Val Ser Asn Leu Ile Tyr Thr Lys Gln Gly Leu Glu Asp Phe 225 230 235 240	720
tgt cag cag tcc gca gtc ata aac aaa gat tgg ttc acg gac ttt gag Cys Gln Gln Ser Ala Val Ile Asn Lys Asp Trp Phe Thr Asp Phe Glu 245 250 255	768
gaa ata cca caa gaa ttg aga att aaa gtt ttt cac ttt gag cat caa Glu Ile Pro Gln Glu Leu Arg Ile Lys Val Phe His Phe Glu His Gln 260 265 270	816
aaa gag atg att agc gca ctc tac gag caa ata gga aga tgt aga gcg Lys Glu Met Ile Ser Ala Leu Tyr Glu Gln Ile Gly Arg Cys Arg Ala 275 280 285	864

cat tta aac gac ttg acc atg aaa atg gca aca cga tta gaa tta ata His Leu Asn Asp Leu Thr Met Lys Met Ala Thr Arg Leu Glu Leu Ile

gca taa

Ala

<210> 14

<211> 305

<212> PRT

<213> Ornithobacterium rhinotracheale

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Gly Lys Leu Met Val Gly Val Lys Pro Pro Leu Thr Pro Asn Gln Glu

Lys Leu Leu Thr Asp Leu Glu Gly Lys Met Glu Ala Gly Thr Ile Thr

Lys Lys Gln Ile Ile Thr Tyr Gly Glu Leu Leu Ser Lys Lys Asn Gln

Lys Leu Glu Leu Ser Ala Ser Val Lys Ser Tyr Leu Ala Asp Ile His

Lys Glu Val Phe Phe Gly Arg Asp Lys Glu Leu Thr Asn Lys Tyr Leu

Ser Lys Gly Ile Gln Val Glu Glu Lys Ser Ile Thr Leu Tyr Ser Asp

Val Cys Asn Lys Leu Phe Leu Lys Asn Lys Lys Phe Tyr Lys Asn Asp

115 120 125

Phe Ile Gln Gly Thr Pro Asp Asn Thr Gln Asp Lys Ile Arg Asp Ile 130 135 140

Lys Ser Ser Trp Asp Phe Ser Thr Phe Pro Leu His Ala Asp Glu Thr 145

Pro Thr Lys Asp Tyr Glu Trp Gln Leu Gln Gly Tyr Met Glu Leu Thr
165 170 175

Gly Leu Lys Glu Ala Glu Leu Ile Tyr Cys Leu Val Asp Thr Pro His 180 185 190

Lys Ile Val Glu Asp Glu Ile Arg Arg Met Asp Trp Lys His Asn Leu 195 200 205

Leu Asp Ile Asn Gly Glu Val Arg Ala Glu Thr Arg Asp Leu Val Val 210 215 220

Glu Ile Val Ser Asn Leu Ile Tyr Thr Lys Gln Gly Leu Glu Asp Phe 225 230 235 240

، دخ

Cys Gln Gln Ser Ala Val Ile Asn Lys Asp Trp Phe Thr Asp Phe Glu 245 250 255

Glu Ile Pro Gln Glu Leu Arg Ile Lys Val Phe His Phe Glu His Gln 260 265 270

Lys Glu Met Ile Ser Ala Leu Tyr Glu Gln Ile Gly Arg Cys Arg Ala 275 280 285

His Leu Asn Asp Leu Thr Met Lys Met Ala Thr Arg Leu Glu Leu Ile 290 295 300 Ala 305 <210> 15 888 <211> DNA <212> <213> Ornithobacterium rhinotracheale <220> <221> CDS <222> (1)..(888) <400> 15 atg aac gaa tta gcg aaa aac gac atc aag tca ttg tta aaa agt gcc 48 Met Asn Glu Leu Ala Lys Asn Asp Ile Lys Ser Leu Leu Lys Ser Ala 15 10 5 1 gac atc aac aaa aga ttt gag caa ttg ctc ggc aaa aaa gca caa ggc 96 Asp Ile Asn Lys Arg Phe Glu Gln Leu Leu Gly Lys Lys Ala Gln Gly 30 25 20 ttt atc tca tca gtc ttg cag acg gca caa aat aac aga ttg tta gcg 144 Phe Ile Ser Ser Val Leu Gln Thr Ala Gln Asn Asn Arg Leu Leu Ala 35 40 aca gcc gac cca aag acc att cta aac gct gca gta aca gcc gcg act 192 Thr Ala Asp Pro Lys Thr Ile Leu Asn Ala Ala Val Thr Ala Ala Thr 60 55 50 tta gat ttg cca att aat cag aat tta ggt tac gcc tac atc gtg cct 240 Leu Asp Leu Pro Ile Asn Gln Asn Leu Gly Tyr Ala Tyr Ile Val Pro 80 75 70 65 tac aaa ggg cag gcg caa ttc caa tta ggc tgg aag ggc ttt gta gca 288 Tyr Lys Gly Gln Ala Gln Phe Gln Leu Gly Trp Lys Gly Phe Val Ala

tta gct aaa aga agt ggc gca tat ttg aaa atg aat gta gta act gtc
Leu Ala Lys Arg Ser Gly Ala Tyr Leu Lys Met Asn Val Val Thr Val
100 105 110

90

85

95

tat caa aat caa ttc aaa tcc tac aat cgc tta aca gaa gaa tta gat Tyr Gln Asn Gln Phe Lys Ser Tyr Asn Arg Leu Thr Glu Glu Leu Asp 115 120 125	384
gcc gat ttc aca atc gaa ggc aat ggt gaa gta gtt ggt tat gca gcc Ala Asp Phe Thr Ile Glu Gly Asn Gly Glu Val Val Gly Tyr Ala Ala 130 135 140	432
tat ttc aaa gaa atc aat ggt ttt gaa aag ctt tcg ttt tgg tca att Tyr Phe Lys Glu Ile Asn Gly Phe Glu Lys Leu Ser Phe Trp Ser Ile 145 150 155 160	480
gag caa gta aaa aaa cac gcc acc aaa tac tct caa act tat ggt aaa Glu Gln Val Lys Lys His Ala Thr Lys Tyr Ser Gln Thr Tyr Gly Lys 165 170 175	528
aaa tca cgc tcg ggg gca tta atg ttt tcg cct tgg aat gat gaa gac Lys Ser Arg Ser Gly Ala Leu Met Phe Ser Pro Trp Asn Asp Glu Asp 180 185 190	576
cag ttt gac gca atg gct atg aag act gtc tta aaa aac acg ctc tca Gln Phe Asp Ala Met Ala Met Lys Thr Val Leu Lys Asn Thr Leu Ser 195 200 205	624
aag ttt ggg aca ctc tca att gaa atg caa atg gcg caa atg gca gac Lys Phe Gly Thr Leu Ser Ile Glu Met Gln Met Ala Gln Met Ala Asp 210 215 220	672
caa gca gtc atc aag aac gag ggg gag tac gag tat ata gac aat acc Gln Ala Val Ile Lys Asn Glu Gly Glu Tyr Glu Tyr Ile Asp Asn Thr 225 230 235 240	720
ata gac att gaa gct gaa agt gcc gaa gaa gaa gcc aat cgt att atg Ile Asp Ile Glu Ala Glu Ser Ala Glu Glu Glu Ala Asn Arg Ile Met 245 250 255	768
aaa ttt att gat aaa gcc gaa agc att gaa gcc tta gag gaa tta aaa Lys Phe Ile Asp Lys Ala Glu Ser Ile Glu Ala Leu Glu Glu Leu Lys 260 265 270	816
tca tca gtt gat gag aat ggc gat tta gag tta tta gcc tat tac gac Ser Ser Val Asp Glu Asn Gly Asp Leu Glu Leu Leu Ala Tyr Tyr Asp 275 280 285	864
aac aga aaa aat gaa tta aaa tga	888

Asn Arg Lys Asn Glu Leu Lys 290 295

<210> 16

÷.

<211> 295

<212> PRT

<213> Ornithobacterium rhinotracheale

<400> 16

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Asp Ile Asn Lys Arg Phe Glu Gln Leu Leu Gly Lys Lys Ala Gln Gly 25 30

Phe Ile Ser Ser Val Leu Gln Thr Ala Gln Asn Asn Arg Leu Leu Ala 35 40 45

Thr Ala Asp Pro Lys Thr Ile Leu Asn Ala Ala Val Thr Ala Ala Thr 50 55 60

Leu Asp Leu Pro Ile Asn Gln Asn Leu Gly Tyr Ala Tyr Ile Val Pro 65 70 75 80

Tyr Lys Gly Gln Ala Gln Phe Gln Leu Gly Trp Lys Gly Phe Val Ala 85 90 95

Leu Ala Lys Arg Ser Gly Ala Tyr Leu Lys Met Asn Val Val Thr Val
100 105 110

Tyr Gln Asn Gln Phe Lys Ser Tyr Asn Arg Leu Thr Glu Glu Leu Asp 115 120 125

Ala Asp Phe Thr Ile Glu Gly Asn Gly Glu Val Val Gly Tyr Ala Ala 130 135 140 Tyr Phe Lys Glu Ile Asn Gly Phe Glu Lys Leu Ser Phe Trp Ser Ile 145 150 155 160

Glu Gln Val Lys Lys His Ala Thr Lys Tyr Ser Gln Thr Tyr Gly Lys
165 170 175

Lys Ser Arg Ser Gly Ala Leu Met Phe Ser Pro Trp Asn Asp Glu Asp 180 185 185

Gln Phe Asp Ala Met Ala Met Lys Thr Val Leu Lys Asn Thr Leu Ser 195 200 205

Lys Phe Gly Thr Leu Ser Ile Glu Met Gln Met Ala Gln Met Ala Asp 210 220

Gln Ala Val Ile Lys Asn Glu Gly Glu Tyr Glu Tyr Ile Asp Asn Thr
225 230 235 240

Ile Asp Ile Glu Ala Glu Ser Ala Glu Glu Glu Ala Asn Arg Ile Met
245 250 255

Lys Phe Ile Asp Lys Ala Glu Ser Ile Glu Ala Leu Glu Glu Leu Lys 260 265 270

Ser Ser Val Asp Glu Asn Gly Asp Leu Glu Leu Leu Ala Tyr Tyr Asp 275 280 285

Asn Arg Lys Asn Glu Leu Lys 290 295